

1 **Decreasing the level of hemicelluloses in sow's lactation diet affects the milk**
2 **composition and post-weaning performance of low birthweight piglets.**

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10 **Abstract**

11 Hemicelluloses (HC) are polysaccharides constituents of the cell walls of plants. They
12 are fermented in the gut to produce volatile fatty acids (VFA). The present study
13 investigated the effects of decreasing HC level in sow's lactation diet on sow
14 performances, offspring development and milk composition. From 110 days (d) of
15 gestation until weaning (26±0.4 d post-farrowing), 40 Swiss Large White sows were
16 assigned to one of the four dietary treatments: (1) T13 (HC: 127g/kg), (2) T11 (HC:
17 114g/kg), (3) T9 (HC: 94g/kg) and (4) T8 (HC: 80g/kg). Milk was collected at 3 and 17d
18 of lactation. At birth, piglets were divided into two groups according to their birthweight
19 (BtW): normal (N-BtW; BtW > 1.20 kg) or low (L-BtW; BtW ≤ 1.20 kg). Decreased HC
20 levels in the maternal diet linearly increased ($P \leq 0.05$) the body weight of L-BtW piglets
21 at two weeks post-weaning and linearly decreased ($P \leq 0.05$) diarrhoea incidence and
22 duration in this category. The concentrations of copper, threonine and VFA, as well as
23 the proportion of butyrate, in milk linearly increased ($P \leq 0.05$), whereas lactose content

24 linearly decreased ($P \leq 0.05$) with decreased HC in the maternal diet. The present
25 study provides evidence that decreasing HC level in sow's lactation diet can positively
26 affect the composition and VFA profile of milk and ultimately favour the growth and
27 health of L-BtW piglets.

28

29 **Keywords:** Dietary fibres, lactose, pigs, volatile fatty acids, butyrate

30

31 **Introduction**

32 Hemicelluloses (HC) represent a complex group of polysaccharides present in the cell
33 walls of all plants, consisting mainly of pentoses (D-xylose and D-arabinose), hexoses
34 (D-galactose, D-glucose and D-mannose) and uronic acids that can be estimated as
35 the difference between NDF and ADF (Huang et al., 2021; Van Soest et al., 1991). As
36 part of dietary fibres (DF), they can resist digestion by endogenous enzymes of the
37 gut. Thus, they can reach the large intestine and promote the growth and activity of
38 beneficial bacteria that produce volatile fatty acids (VFA) (Lattimer and Haub, 2010).
39 These latter, namely acetate, propionate and butyrate, provide up to 28% of the energy
40 requirements in growing pigs and even more in sows, where they can be absorbed and
41 transferred to the milk and serve as an energy source for milk synthesis (Noblet and
42 Le Goff, 2001; Tian et al., 2020). A previous study focused on increasing the level of
43 DF in sow's gestation diet showed that adding up to 20% DF increases colostrum fat
44 content, as well as colostrum intake, of low birthweight (BtW) piglets ($0.6 \text{ kg} \leq \text{BtW}$
45 $< 0.9 \text{ kg}$) and decreases litter mortality during the suckling period (Loisel et al., 2013).
46 ~~However, this aspect can be relevant, since inclusion of DF during lactation may also~~
47 ~~positively affect the development and gut health of piglets.~~ These positive effects on

48 growth and survival are in line with findings of Paßlack et al. (2015) who reported that
49 inclusion of 3% inulin, a source of DF offered to lactating sows positively affected the
50 development and gut health of their litters.

51 Like all DF, HC can be differentiated according to their physiochemical properties such
52 as their solubility in water and their intestinal fermentability. Apart from the quantity of
53 DF provided, the beneficial effects of DF is also related to their physiochemical
54 properties such as their solubility in water and their intestinal fermentability. For
55 instance, due to a slower fermentability compared to soluble dietary fibres (SDF), the
56 majority of the insoluble dietary fibres (IDF) reach the large intestine and stimulate the
57 growth of commensal and probiotic bacteria such as *Ruminococcus*, *Faecalibacterium*,
58 *Lactobacillus* and *Bifidobacterium* (LeBlanc et al, 2017). The large intestine acts then
59 as a fermentation chamber producing VFA, CO₂, H₂ and other carboxylic acids
60 (Lattimer and Haub, 2010). Conversely, compared to insoluble dietary fibres (IDF),
61 SDF are easily fermented and may be completely degraded at the end of the small
62 intestine (Houdijk et al., 2002). Depending on the plants, HC might be considered as a
63 source of SDF (Jiménez-Escrig et al., 2000). A previous study in growing pigs reported
64 that decreasing HC level increased VFA produced in the ileum (Zhao et al., 2019).
65 However, to our knowledge, little is known about the effects of HC level in lactating
66 sows. Therefore, the present study aims to fill this gap by comparing four diets
67 characterised by similar total DF and crude fibre content but different HC levels by
68 varying the sources of DF. We hypothesised that decreasing the level of HC while
69 maintaining a similar total DF level in sow's lactation diet would may affect the IDF to
70 SDF ratio and by that impact gut fermentation particularly in the large intestine and
71 ultimately modify milk composition.

72 **Materials and methods**

73 ***Animals, housing and treatments***

74 The experiment was conducted during late gestation and lactation of 40 Swiss Large
75 White sows from five farrowing batches. Approximately 10 days before the expected
76 time of farrowing, sows were moved to farrowing rooms arranged with individual 7.1
77 m² farrowing crates, consisting of a 5.89 m² concrete solid floor and a 1.21 m² concrete
78 slatted floor. Each crate was equipped with an electronic sow feeder (Schauer Spotmix,
79 Schauer Agrotronic GmbH, Austria), nipple drinker and a heated covered area for
80 piglets. The ambient temperature was maintained at 24 °C, and artificial lights were on
81 from 0800 h to 1700 h. On day 110 of gestation, the sows were randomly allocated to
82 one of the four experimental lactation diets based on parity (mean ± SEM: 3.5 ± 0.7)
83 and BW (mean ± SEM: 286.5 ± 13.6 kg). Parturition was induced when gestation period
84 exceeded 116 days with an intramuscular injection of 1 ml (0.25 g/ml) of cloprostenol
85 (Estrumate®, MSD Animal Health GmbH, Luzern, Switzerland). Within the first 24 h
86 following birth, piglets were identified by an individual ear tag and received iron
87 injection (Feridex® 10%, AMAG Pharmaceuticals, Inc., Waltham, USA). Piglets
88 weighing less than 800 g at birth were excluded from the experiment. To adjust litter
89 size to an average of 12 piglets per sow, cross-fostering was carried out only on male
90 piglets 24 h post-farrowing. After anaesthetisation, the male piglets were castrated in
91 the second week. Piglets were weaned on day 25.7 ± 0.44 (mean ± SEM) of age but
92 were kept in their respective farrowing crates until 2 weeks post-weaning. The heating
93 nest temperature was set at 40 °C following birth and then gradually decreased by 0.5
94 °C per day to reach a final temperature of 32 °C

95 ***Diets and feeding***

96 The experimental diets were formulated to be isonitrogenous and isocaloric (Table 1)
 97 and to differ in DF sources and HC content: (1) T13 (HC: 127 g/kg), (2) T11 (HC: 114
 98 g/kg), (3) T9 (HC: 94 g/kg) and (4) T8 (HC: 80g/kg). The daily feed allowance was
 99 calculated according to the current Swiss feeding recommendations for pigs
 100 (Agroscope, 2018). Sows had *ad libitum* access to water and were provided with
 101 moderate quantities of straw bedding, as required by the Swiss legislation. During the
 102 end of gestation, feed allowance was on average 3.04 ± 0.16 kg (mean \pm SEM). While,
 103 during lactation, the feed allowance was gradually increased by 0.5 kg/day until *ad*
 104 *libitum* feeding on day 12 of lactation approximatively. All diets were delivered three
 105 times per day in three equal meals using a computerised feed delivery system
 106 (Schauer Spotmix, Schauer Agrotronic GmbH, Austria). Throughout the experiment,
 107 the feed refusals of the sows were weighed daily to calculate actual feed intake. From
 108 day 18.7 ± 0.44 of age (mean \pm SEM) to 2 weeks post-weaning (mean \pm SEM: day
 109 39.7 ± 0.44 of age), piglets had *ad libitum* access to a post-weaning standard starter
 110 diet and water. The post-weaning starter diet contained 170 g/kg crude protein, 58 g/kg
 111 fat, 50 g/kg crude fibre and 14 MJ/kg digestible energy.

112 **Table 1. Ingredients and composition of the sow's lactation diet**

Item	Dietary Treatments ¹			
	T13	T11	T9	T8
Ingredients (%)				
Barley, ground	54.4	38.7		4.7
Oat flakes			4.0	18.2
Corn, ground	10.3		26.9	16.0
Rye		25.0	10.0	
Wheat, ground			13.1	15.0
Wheat starch	4.0	4.0	4.0	4.0
Molasses				4.0
Animal fat RS 65	2.4	2.4	3.0	3.8
Potato protein	10.0	10.0	10.0	10.0

Soybean meal	10.0	10.0	10.0	10.0
Flaxseed Meal	0.6			
Rapeseed meal		0.4		1.7
Oat hulls			4.0	8.0
Lupin			2.5	
Wheat bran			4.0	
Beet pulp	3.0	5.0	4.0	
L-lysine-HCL	0.070	0.057	0.057	0.056
DL-methionine	0.200			
L-threonine	0.050			0.050
L-tryptophan	0.020	0.006	0.013	0.003
Dicalcium phosphate	0.94	0.70	0.82	0.85
Calcium carbonate	1.57	1.38	1.39	1.47
Salt	0.59	0.52	0.42	0.41
Pellan ²	0.40	0.40	0.40	0.40
Celite	1.00	1.00	1.00	1.00
Premix ³	0.40	0.40	0.40	0.40
Natuphos 5000 G ⁴	0.01	0.01	0.01	0.01
Gross chemical composition analysed (g/kg as fed)				
Dry matter	900	894	897	900
Crude protein	193	191	192	196
Fat	51	46	57	60
Crude fibre	43	43	47	46
Ash	63	61	60	63
NDF	184	174	163	154
ADF	57	60	69	79
Hemicelluloses ⁵	127	114	94	80
Total dietary fibres	210	227	220	203
Low-molecular-weight dietary fibres	18	23	18	14
Soluble dietary fibres	43	44	35	28
Insoluble dietary fibres	149	160	167	161
IDF/SDF ⁶	3.46	3.63	4.77	5.75
Calcium	9.4	9.4	9.3	8.7
Phosphorus	5.0	4.6	5.0	4.7
Gross chemical composition calculated				
Digestible energy (MJ/kg)	14.1	14.1	14.1	14.1
Digestible phosphorus (g/kg as fed)	3.1	2.8	2.8	2.8
Digestible essential amino acids (g/kg as fed)				
Lysine	9.6	9.6	9.6	9.6
Methionine	4.9	2.9	3.0	3.0

Threonine	6.9	6.3	6.4	6.9
Tryptophan	2.0	1.8	1.8	1.8

113 ¹T13= Sow's lactation diet containing 13% of hemicelluloses; T11 = Sow's lactation diet containing 11%
114 of hemicelluloses; T9 = Sow's lactation diet containing 9% of hemicelluloses; T8 = Sow's lactation diet
115 containing 8% of hemicelluloses.

116 ²Pellet binding aid: Pellan, Mikro-Technik, Bürgstadt, Germany.

117 ³Supplied per kg of diet: vitamin A, 8000 IU; vitamin D3, 800 IU; vitamin E, 40 mg; menadione, 2 mg;
118 thiamine, 2 mg; riboflavin, 5 mg; biotin, 0.1 mg; niacin, 20 mg; pantothenic acid, 20 mg; iodine (as
119 calcium iodate), 0.55 mg; copper (as copper sulphate), 7 mg; manganese (as manganese oxide), 20
120 mg; zinc (as zinc oxide), 55 mg; selenium (as sodium selenite), 0.2 mg.

121 ⁴Phytase supplemented with 500 units of *Aspergillus niger* phytase/kg diet.

122 ⁵Hemicellulose: calculated as the difference between NDF and ADF.

123 ⁶Ratio of insoluble to soluble dietary fibres

124 ***Sow and piglet performance***

125 The BW of the sows, body condition score (BCS) and backfat thickness were recorded
126 at the 110th day of gestation and on the day of farrowing and weaning. Weight loss
127 during lactation was calculated as the weight difference between farrowing and
128 weaning. Based on visual observation and palpations, BCS was determined according
129 to a scale ranging from 1 (*very thin*) to 6 (*obese*) points (Dourmad et al., 2001),
130 including intermediate values of 0.33 points. Briefly, the trained personnel assessed
131 sows by palpating the shoulders, ribs, backbone and hips, followed by a visual
132 observation. Backfat thickness was measured on each side at 65 mm of the dorsal
133 midline at the level of the last rib (P2) using a digital ultrasound back-fat indicator
134 (Renco Lean Meter Digital Backfat Indicator, Renco Corporation, Minneapolis,
135 Minnesota, USA). Backfat thickness loss during lactation was then calculated as the
136 difference between backfat thickness measurements during farrowing and weaning. At
137 farrowing, the number of born alive, stillborn and mummified piglets were recorded
138 within each litter. Farrowing was recorded using a digital video recorder to estimate the
139 farrowing duration, which is defined as the time span between the time of birth of the

140 first and last piglet of the litter. At birth, the piglets were individually weighed, and
141 crown-to-rump length and body circumference were recorded. Piglets were then
142 individually weighed 5 and 16 days postpartum, during weaning (mean \pm SEM: 25.7 \pm
143 0.44 days of age) and at 1 (mean \pm SEM: 32.7 \pm 0.44 days of age) and 2 weeks post-
144 weaning (mean \pm SEM: 39.7 \pm 0.44 days of age). The average daily gain (ADG) and
145 litter weight during birth and weaning were calculated from these data. Milk yield was
146 calculated as the individual piglet gain summed in the same litter multiplied by a
147 numerical coefficient of 4.2 (Van der Peet-Schwering et al., 1998). The indices of body
148 conformation were calculated based on the measurements of the individual BtW and
149 the crown-to-rump length. The body mass index was calculated as the ratio of BtW to
150 the squared value of the crown-to-rump length, and the ponderal index was calculated
151 as the ratio of BtW to the cubic value of the crown-to-rump length (Hales et al., 2013).
152 In addition, piglets were divided into two BtW groups: normal (N-BtW; BtW > 1.20 kg)
153 or low (L-BtW; BtW \leq 1.20 kg). From 1 week before weaning onwards, feed intake and
154 refusals (including feed waste) per pen as well as the occurrence of diarrhoea were
155 recorded daily. Diarrhoea incidence was determined according to a daily faecal score
156 assessed using a scale from 0 = *no diarrhoea* to 1 = *diarrhoea*. The percentage of
157 diarrhoea per group was calculated as the sum of piglets with a faecal score of one
158 divided by the total number of piglets.

159 **Sample Collection**

160 Within each farrowing series, feed samples of the four diets were collected weekly and
161 pooled over the experimental period to determine the chemical composition. On days
162 3 and 17 of lactation, milk samples were manually collected from all functional teats
163 after an intramuscular injection of 2 ml of oxytocin (Intertocine-S, MSD Animal Health

164 GmbH, Luzern, Switzerland). Before milking, the piglets were temporarily isolated from
165 the sow for 2 h, and the teats were cleaned with humid wipes. One aliquot of milk was
166 refrigerated at 5 °C with 4 mg of bronopol to determine somatic cell concentration, and
167 three aliquots were immediately stored at –20 °C for further analysis.

168 ***Analytical Methods***

169 *Feed Analysis*

170 After being ground to pass a 1-mm screen (Brabender rotary mill; Brabender GmbH &
171 Co. KG, Duisburg, Germany), feed samples were analysed for dry matter content by
172 heating at 105°C for 3h followed by incineration at 550°C until a stable mass was
173 reached to determine the ash content according to ISO 5984:2002 (prepASH, Precisa
174 Gravimetrics AG, Dietikon, Switzerland). An inductively coupled plasma optical
175 emission spectrometer (ICP-OES, Optima 7300 DV; Perkin-Elmer, Schwerzenbach,
176 Switzerland) was used to measure mineral content (European Standard EN
177 15510:2008). The CP content was calculated as nitrogen (N) content multiplied by a
178 coefficient of 6.25, where N was determined with the Dumas method (ISO 16634–
179 1:2008). Fat content was extracted with petrol ether after acid hydrolysis (ISO
180 6492:1999). Different categories of fibres were analysed by standard protocols. Crude
181 fibre content was determined gravimetrically (ISO 6865:2000) by incineration of
182 residual ash after acid and alkaline digestions using a fibre analyser (Fibretherm
183 Gerhardt FT-12, C. Gerhardt GmbH & Co. KG, Königswinter, Germany). The NDF and
184 ADF contents (ISO 16472:2006 for NDF and ISO 13906:2008 for ADF) were analysed
185 with the same fibre analyser (Fibretherm Gerhardt FT-12, C. Gerhardt GmbH & Co.
186 KG, Königswinter, Germany) and were expressed without residual ash. NDF
187 determination was evaluated with heat stable amylase and sodium sulfite and

188 expressed without residual ash after incineration at 600°C for 3 h. The contents of SDF,
189 IDF and low-molecular-weight DF were measured according to AOAC Method
190 2011.25, and the total DF content was calculated as the sum of the three
191 aforementioned types of DFs.

192 *Milk Analysis*

193 The dry matter of the frozen milk samples was determined after freeze-drying (Christ
194 DELTA 2-24 LSC, Kühner AG, Birsfelden, Switzerland) for 70 hours. Subsequently,
195 freeze-dried samples were milled with a mortar. Residual dry matter, ash, mineral and
196 nitrogen contents were analysed as previously described for the feed chemical
197 analysis, except that CP was expressed as N x 6.38. Except for tryptophan, all amino
198 acids were determined as described in ISO 13903:2005. Briefly, after oxidation, 24 h
199 of acid hydrolysis occurred with 6M HCl and derivatization with AccQ-Tag Ultra reagent
200 (Waters corporation, Milford, USA USA), the amino acid profile was determined by
201 ultra-high-performance liquid chromatography (UHPLC) coupled with a UV detector
202 (Vanquish, Thermo Scientific, Reinach, Switzerland. Tryptophan content was
203 quantified by HPLC (LC 1290 Infinity II LC System, Agilent Technologies, USA)
204 according to ISO 13904:2016. Gross energy content was determinate by combustion
205 in a calorimetric vessel under pure oxygen condition using an adiabatic bomb
206 calorimeter (AC600 Semi-Automatic Calorimeter, Leco Corporation, USA) (ISO
207 9831:1998). Lactose content was determined by enzymatic testing with β -
208 galactosidase and galactose dehydrogenase (Enzytec TM Liquid Lactose/D-Galactose
209 Ref. No. E8110, R-Biopharm AG, Darmstadt, Germany). Somatic cells count (ISO
210 13366-2) was determined by flow cytometry (Somacount FC, Bentley Instruments Inc.,
211 USA). Fatty acid methyl esters, as described by Kragten et al. (2014), and the VFA

212 profile (ISO 15884:2002) (ISO 15885:2002) were determined by gas-liquid
213 chromatography (Gaschromatograph Series II Agilent 6850, Agilent Technologies
214 2000, USA and Gaschromatograph Serie Agilent 6890, Agilent Technologies 2000,
215 USA, respectively). Fat content was determined as total fatty acids multiplied by a
216 coefficient of 1.05.

217 **Statistical Analysis**

218 Due to health problems that could not be related to the dietary treatment, one T9 sow
219 was excluded from the experiment. Data were analysed by ANOVA using the 'lme' and
220 the 'glmmPQL' function of the *nlme* package of R Studio (version 4.0.2 for Windows).
221 Regarding sow performance, milk composition and VFA profile, the sow was the
222 experimental unit; the pen was the experimental unit regarding piglet feed intake and
223 litter performance; and the piglet was the experimental unit of piglet's individual
224 performance, days and percentage of diarrhoea. Linear regression models, including
225 the treatment and the farrowing batch as fixed effects, were used to fit data related to
226 sow performance, litter performance, piglet feed intake and days with diarrhoea. Data
227 related to piglets' individual performance were analysed using a linear mixed-effects
228 model, including the treatment and the farrowing batch as fixed effects and the sow as
229 random effects. Milk composition and VFA profile were analysed with a linear mixed-
230 effects model and fitted in repeated measurements, including the treatment, the
231 farrowing batch, the sampling day, and the interaction between the treatment and
232 sampling day as fixed effects and the sow as random effect. Before analysis,
233 logarithmic transformation was applied to the milk fatty acid and milk VFA data due to
234 the non-normality of the residuals. The percentage of diarrhoea was analysed using a
235 generalised linear mixed model using Penalized Quasi-Likelihood, including the

236 treatment, the farrowing batch and the day as fixed effects and the piglet as a random
237 factor. Orthogonal polynomial contrasts were implemented to evaluate the linear or
238 quadratic effects of decreasing HC level. The results are expressed as the least square
239 means \pm SEM. Linear and quadratic effects were considered significant at $P \leq 0.05$.

240 **Results**

241 ***Sows' performance***

242 The sow BW, BCS and backfat thickness on day 110 of gestation and during farrowing
243 and weaning were not influenced by the dietary treatment, resulting in similar weight
244 and backfat thickness losses during the lactation period. Daily feed intake in the pre-
245 farrowing period and during lactation did not differ between treatments. Fibre intake
246 was partially influenced by dietary treatments. In both the pre-farrowing and lactation
247 periods, the NDF, HC, (linear effects; $P < 0.01$), low-molecular-weight DF and SDF
248 intake decreased (linear and quadratic effects; $P < 0.01$), and the ADF intake increased
249 (linear effect; $P < 0.01$) with decreasing HC levels in the diet. A quadratic effect ($P \leq$
250 0.04) of the HC level was found in the diets on the intake of total DFs in the pre-
251 farrowing and lactation periods. At birth, litter traits, such as total born, born alive and
252 stillborn piglets, did not differ, leading to comparable litter weights in the four
253 treatments. Likewise, the dietary treatments had no effect on the total number of piglets
254 weaned and, consequently, on litter weight at weaning. Farrowing duration was not
255 influenced by dietary treatments. During the entire lactation period, milk yield was not
256 influenced by the dietary treatments, with an average estimated production of 10.38
257 kg/day per sow (Table 2).

258 **Table 2. Effect of decreasing hemicelluloses level in lactation diet on sow's**
259 **performance**

Item	¹ Dietary Treatments				SEM	² Contrasts	
	T13	T11	T9	T8		L	Q
Sows							
Number of sows, <i>n</i>	10	10	9	10			
Range of parity, <i>n</i>	3.8	3.8	3.5	3.5	0.69	0.51	0.99
Farrowing duration, min	308	337	321	262	70.7	0.54	0.44
Body weight, kg							
D110	284	291	284	287	13.6	0.71	0.89
Farrowing	264	267	269	272	14.1	0.67	0.99
Weaning	233	238	248	246	12.5	0.39	0.78
Weight loss in lactation, kg	30.6	28.7	20.9	26.1	2.58	0.19	0.30
BCS, <i>n</i>							
D110	4.09	4.10	4.03	3.83	0.129	0.79	0.18
Farrowing	3.58	3.59	3.40	3.64	0.148	0.96	0.40
Weaning	2.71	2.62	2.81	2.94	0.246	0.42	0.62
Backfat thickness, mm							
D110	13.8	14.8	12.7	15.8	0.88	0.31	0.23
Farrowing	13.7	14.6	12.7	15.6	0.88	0.34	0.24
Weaning	11.3	11.9	11.5	12.9	0.71	0.18	0.52
Backfat thickness loss in lactation, mm	2.38	2.66	1.25	2.67	0.505	0.82	0.24
Milk yield, kg/day	10.61	10.85	10.09	9.97	0.720	0.41	0.79
Feed intake, kg/day							
Pre-farrowing	2.93	3.03	3.03	3.00	0.155	0.75	0.68
Lactation	5.67	5.93	5.77	5.87	0.237	0.69	0.73
Fibre intake, g/day							
Pre-farrowing							
Crude fibre	127	129	141	139	7.0	0.13	0.74
NDF	538	527	492	461	26.1	0.03	0.69
ADF	168	182	208	224	10.3	<0.01	0.90
Hemicelluloses	370	345	284	237	16.1	<0.01	0.48
Total dietary fibres	614	688	667	610	33.1	0.82	0.04
Low-molecular-weight dietary fibre	53	70	55	42	2.8	<0.01	<0.01
Soluble dietary fibres	133	126	106	83	5.8	<0.01	<0.01
Insoluble dietary fibres	436	485	506	484	24.8	0.14	0.14
Lactation							
Crude fibre	246	254	269	272	10.4	0.06	0.81
NDF	1043	1033	937	903	41.3	<0.01	0.75
ADF	325	356	396	438	14.8	<0.01	0.71
Hemicelluloses	718	677	541	465	26.8	<0.01	0.49
Total dietary fibres	1190	1350	1270	1190	51.1	0.76	0.02
Low-molecular-weight dietary fibre	102	137	104	82	4.6	<0.01	<0.01
Soluble dietary fibres	244	261	202	163	9.6	<0.01	<0.01
Insoluble dietary fibres	845	950	964	947	37.1	0.06	0.09
Suckling piglets							
Number of piglets per litter, <i>n</i>							
Total born ³	13.5	13.7	13.5	14.3	1.12	0.65	0.76

Born alive ³	12.8	12.4	11.4	12.7	1.27	0.82	0.49
Stillborn	0.7	1.3	2.1	1.6	0.65	0.22	0.40
After cross-fostering	11.4	11.4	11.5	11.6	0.79	0.85	0.91
Weaned	10.7	10.9	11.3	10.7	0.76	0.94	0.60
Litter weight, kg							
At birth	20.5	20.5	21.1	20.3	1.63	0.99	0.81
At weaning	81.9	83.9	78.0	79.4	5.50	0.59	0.96

260 ¹T13 = Sow's lactation diet containing 13% of hemicelluloses; T11 = Sow's lactation diet containing 11%
261 of hemicelluloses; T9 = Sow's lactation diet containing 9% of hemicelluloses; T8 = Sow's lactation diet
262 containing 8% of hemicelluloses.

263 ²Contrasts: L = Linear; Q = Quadratic.

264 ³ including piglets weighing less than 800g at birth.

265 ***Piglets' individual performance***

266 Body characteristics, such as body circumference, crown-to-rump length, body mass
267 index and ponderal index, were not affected by the lactation diet of the sows (Table 3).
268 Similarly, piglet BW development, ADG and feed intake were not affected by the dietary
269 treatments. During the first week post-weaning, the incidence of diarrhoea and the
270 number of days with diarrhoea were similar among the treatments. By contrast, during
271 the second week of post-weaning, a quadratic increase ($P \leq 0.05$) in the incidence of
272 diarrhoea and the number of days with diarrhoea was observed with decreasing HC
273 level. When focusing on the two BtW categories, the effect of the sow diets in the L-
274 BtW group showed interesting observations (Table 4). The BtW, the BWs until one
275 week post-weaning and in accordance the ADG in this period were similar among the
276 experimental treatments for L-BtW piglets. By contrast, the decrease in HC level in the
277 sow diets increased (linear effect; $P \leq 0.04$) the BW and the ADG in the second week
278 post-weaning and the overall ADG from birth to two weeks post-weaning of L-BtW
279 piglets. In the first week post-weaning, the dietary treatments did not affect either the
280 incidence of diarrhoea or the days with diarrhoea of L-BtW piglets. In the second week
281 post-weaning, the incidence of diarrhoea and days with diarrhoea linearly decreased

282 (P < 0.01) with decreased HC level in the maternal diet. Except for the linear increase
283 in the incidence of diarrhoea and increase in the number of days with diarrhoea in the
284 second week post-weaning (P < 0.01) with decreasing HC level, no dietary effects on
285 growth traits were observed in N-BtW pigs (Supplementary Table 1).

286 **Table 3. Effect of decreasing hemicelluloses level in the maternal diet on the**
 287 **performance of piglets**

	¹ Dietary Treatments				SEM	² Contrasts	
	T13	T11	T9	T8		L	Q
Body measurements at birth, cm							
Crown-to-rump length	28.7	28.9	28.8	28.4	0.53	0.60	0.56
Body circumference	25.5	25.6	25.8	25.3	0.49	0.81	0.57
Body mass index, kg/m ²	19.2	18.8	19.5	18.7	0.55	0.78	0.73
Ponderal index, kg/m ³	67.2	65.1	67.9	66.3	2.03	0.99	0.88
Body weight, kg							
At birth	1.61	1.60	1.63	1.52	0.083	0.55	0.50
5 days post-farrowing	2.38	2.37	2.45	2.22	0.126	0.49	0.36
16 days post-farrowing	5.36	5.28	5.25	4.94	0.272	0.29	0.65
Weaning	7.69	7.54	7.26	7.36	0.348	0.41	0.71
1 week post-weaning	7.82	7.69	7.42	7.48	0.371	0.55	0.92
2 week post-weaning	8.93	9.17	8.71	8.93	0.453	0.82	0.99
ADG, g/day							
Birth to 5 days post-farrowing	154	154	160	137	12.6	0.44	0.34
Birth to 16 days post-farrowing	235	230	225	212	13.9	0.25	0.74
Birth to weaning	237	232	222	222	11.5	0.29	0.82
Weaning to 2 weeks post-weaning	86	116	103	113	17.3	0.38	0.52
1 week to 2 weeks post-weaning	172	194	184	207	20.8	0.31	0.98
Birth-2 week post-weaning	185	191	180	184	9.9	0.73	0.90
Feed intake, g/piglet							
1 week pre-weaning	182	186	157	189	24.5	0.95	0.55
1 week post-weaning	753	883	760	786	121.0	0.96	0.65
2 weeks post-weaning	1428	1638	1436	1600	144.0	0.63	0.87
Post-weaning diarrhoea, %							
1 week post-weaning	26.1	29.3	27.0	29.6	2.47	0.47	0.77
2 weeks post-weaning	17.4	17.2	12.8	22.2	2.82	0.44	0.05
Days with diarrhoea, days							
1 week post-weaning	1.89	2.09	1.85	2.10	0.171	0.61	0.90
2 weeks post-weaning	1.45	1.40	1.11	1.80	0.172	0.34	0.02

288 ¹T13= Sow's lactation diet containing 13% of hemicelluloses; T11 = Sow's lactation diet containing
 289 11% of hemicelluloses; T9 = Sow's lactation diet containing 9% of hemicelluloses; T8 = Sow's
 290 lactation diet containing 8% of hemicelluloses.
 291 ²Contrasts: L = Linear; Q = Quadratic.

292 **Table 4. Effect of decreasing hemicelluloses level in maternal diet on the**
 293 **performance of low birthweight piglets**

	¹ Dietary Treatments				SEM	² Contrasts	
	T13	T11	T9	T8		L	Q
Number of piglets, <i>n</i>	25	23	15	20			
Body measurements at birth, cm							
Crown-to-rump length	25.0	25.4	25.5	26.0	0.56	0.22	0.97
Body circumference	22.3	21.8	21.9	22.3	0.42	0.98	0.16
Body mass index, kg/m ²	16.6	15.5	16.1	15.8	0.65	0.51	0.43
Ponderal index, kg/m ³	66.6	61.4	63.6	61.7	3.22	0.37	0.54
Body weight, kg							
At birth	1.04	1.01	1.04	1.06	0.047	0.64	0.46
5 days post-farrowing	1.61	1.59	1.58	1.54	0.095	0.53	0.86
16 days post-farrowing	3.94	3.78	3.59	3.85	0.287	0.70	0.39
Weaning	5.86	5.73	5.42	6.55	0.468	0.38	0.12
1 week post-weaning	5.92	5.96	5.56	6.95	0.498	0.20	0.12
2 week post-weaning	6.55	6.66	6.43	8.35	0.545	0.02	0.06
ADG, g/day							
Birth to 5 days post-farrowing	113	118	104	91	14.1	0.14	0.43
Birth to 16 days post-farrowing	181	173	158	173	16.7	0.57	0.41
Birth to weaning	192	184	171	201	16.3	0.83	0.18
Weaning to 2 weeks post-weaning	50	62	74	113	27.0	0.09	0.56
1 week to 2 weeks post-weaning	91	103	125	187	27.0	0.01	0.25
Birth to 2 weeks post-weaning	141	143	135	177	11.5	0.04	0.05
Post-weaning diarrhoea, %							
1 week post-weaning	19.8	34.1	16.4	20.8	10.50	0.69	0.50
2 weeks post-weaning	36.4	16.7	6.5	5.2	8.31	<0.01	0.35
Days in diarrhoea, days							
1 week post-weaning	1.66	2.26	1.32	1.63	0.502	0.56	0.71
2 weeks post-weaning	2.36	1.22	0.55	0.87	0.512	<0.01	0.07

294 ¹T13 = Sow's lactation diet containing 13% of hemicelluloses; T11 = Sow's lactation diet containing
 295 11% of hemicelluloses; T9 = Sow's lactation diet containing 9% of hemicelluloses; T8 = Sow's
 296 lactation diet containing 8% hemicellulose.

297 ²Contrasts: L = Linear; Q = Quadratic.

298 **Milk Composition**

299 Throughout lactation, no dietary treatment and sampling day interaction was found
300 (data not shown). At days 3 and 17 of lactation, DM, ash, protein and somatic cell
301 count, as well as milk yield estimated from farrowing to day 3 and from day 4 to day 17
302 of lactation, were similar among dietary treatments (Table 5). With a decreasing HC
303 level, milk lactose content linearly decreased ($P < 0.01$). Regarding mineral levels in
304 the sow milk, calcium, phosphorus, sodium, magnesium and zinc contents remained
305 similar among experimental treatments, whereas the copper content linearly increased
306 ($P = 0.02$) with decreasing HC content in the maternal diet. Excluding the linear
307 increase ($P = 0.04$) in the threonine level and the quadratic increase ($P = 0.04$) in the
308 monounsaturated fatty acid portion, decreasing HC level in the maternal diet had no
309 impact on the amino acid and fatty acid profiles. Regardless of the dietary treatments,
310 somatic cell counts did not differ between the sampling days. However, the sampling
311 day influenced protein, mineral and lactose contents, as well as milk yield. Between
312 days 3 and 17 of lactation, protein, phosphorus, potassium and zinc contents
313 decreased ($P \leq 0.05$), whereas lactose and calcium contents and milk yield increased
314 ($P \leq 0.05$). Furthermore, histidine, leucine, isoleucine, phenylalanine, threonine,
315 tryptophan, tyrosine, valine, alanine, aspartic acid and serine decreased ($P \leq 0.05$),
316 whereas glutamate and proline increased ($P \leq 0.05$) between days 3 and 17. The fatty
317 acid profile in milk changed during lactation. Monounsaturated and polyunsaturated
318 fatty acid portions decreased ($P \leq 0.05$) and saturated fatty acid content increased (P
319 ≤ 0.05) from day 3 to day 17. More precisely, the portions of C18:0, C18:1n-9, C18:2n-
320 6, C18:3n-6, C18:3n-3, C20:4n-6, C20:5n-3 and C22:5n-3 decreased ($P \leq 0.05$),
321 whereas C16:0 level increased ($P \leq 0.05$) between days 3 and 17.

322 **Table 5. Effect of decreasing hemicellulose level in sow's lactation diet on gross composition, mineral content, amino acid**
 323 **profile and fatty acid profile of milk**

Item	¹ Dietary Treatments				SEM	² Contrasts		³ Stage of lactation		SEM	<i>P</i> -value
	T13	T11	T9	T8		<i>L</i>	<i>Q</i>	d3	d17		
Milk yield, kg/day	9.66	9.90	9.39	8.80	0.76	0.81	0.76	7.03	11.85	0.41	<0.01
Gross chemical composition											
Dry matter, %	19.5	20.7	19.9	20.6	0.60	0.25	0.49	20.7	19.7	0.40	0.06
Total protein, %	5.86	5.82	5.84	6.07	0.153	0.48	0.34	6.40	5.40	0.091	<0.01
Fat, %	7.50	8.65	8.07	8.69	0.533	0.15	0.38	8.50	7.96	0.364	0.27
Lactose, %	5.17	4.99	4.92	4.77	0.110	0.01	0.76	4.56	5.37	0.068	<0.01
Ash, %	0.86	0.86	0.88	0.85	0.150	0.29	0.89	0.89	0.83	0.098	<0.01
Somatic cells, log 10 ³ cells/ml	6.99	6.92	7.40	7.71	0.325	0.18	0.41	7.40	7.11	0.248	0.93
Gross energy, MJ/kg	5.14	5.70	5.43	5.70	0.230	0.10	0.30	5.65	5.34	0.162	0.72
Minerals											
Calcium, g/kg	1.91	1.98	2.02	1.99	0.051	0.97	0.94	1.88	2.07	0.033	<0.01
Phosphorus, g/kg	1.57	1.58	1.57	1.53	0.026	0.09	0.63	1.61	1.52	0.017	<0.01
Potassium, g/kg	1.11	1.10	1.11	1.05	0.028	0.07	0.24	1.29	0.90	0.019	<0.01
Sodium, g/kg	0.37	0.35	0.35	0.34	0.016	0.21	0.89	0.36	0.34	0.011	0.93
Magnesium, g/kg	0.10	0.11	0.11	0.11	0.003	0.42	0.09	0.11	0.11	0.002	0.57
Copper, mg/kg	1.37	1.45	1.51	1.76	0.135	0.02	0.28	1.68	1.37	0.085	0.67
Zinc, mg/kg	6.04	6.64	6.02	5.44	0.363	0.16	0.07	6.38	5.69	0.213	<0.01
Amino acids, % of total protein											
Alanine	3.28	3.29	3.33	3.35	0.025	0.21	0.46	3.41	3.21	0.017	<0.01
Arginine	4.57	4.62	4.68	4.67	0.029	0.13	0.99	4.72	4.55	0.020	<0.01
Aspartic acid	7.70	7.68	7.75	7.74	0.035	0.78	0.29	7.83	7.61	0.025	<0.01
Cysteine	1.40	1.39	1.39	1.42	0.015	0.15	0.10	1.44	1.36	0.010	<0.01
Glutamate	17.8	17.6	17.8	17.6	0.16	0.29	0.99	17.5	17.9	0.11	<0.01
Glycine	2.98	3.02	3.12	3.05	0.030	0.10	0.40	3.06	3.03	0.019	0.15

Histidine	2.53	2.53	2.53	2.56	0.015	0.85	0.31	2.56	2.51	0.009	<0.01
Isoleucine	3.85	3.80	3.80	3.83	0.039	0.36	0.53	3.84	3.80	0.022	0.05
Leucine	8.03	8.12	8.02	8.15	0.049	0.74	0.83	8.18	7.99	0.030	<0.01
Lysine	6.86	6.79	6.82	6.86	0.049	0.61	0.36	6.85	6.82	0.029	0.22
Methionine	1.74	1.72	1.71	1.71	0.014	0.10	0.68	1.72	1.72	0.008	0.52
Phenylalanine	3.86	3.85	3.87	3.92	0.026	0.17	0.13	3.92	3.83	0.017	<0.01
Proline	10.2	10.3	10.4	10.2	0.11	0.37	0.15	10.1	10.5	0.07	<0.01
Serine	4.70	4.66	4.73	4.76	0.047	0.15	0.39	4.75	4.67	0.030	0.02
Threonine	3.88	3.88	3.90	3.98	0.036	0.04	0.19	3.99	3.83	0.023	<0.01
Tryptophan	1.18	1.18	1.21	1.20	0.017	0.17	0.94	1.23	1.15	0.011	<0.01
Tyrosine	4.02	3.97	3.99	4.05	0.050	0.43	0.20	4.05	3.96	0.028	<0.01
Valine	5.16	5.21	5.20	5.26	0.039	0.17	0.98	5.30	5.12	0.025	<0.01
Fatty acids, % of total fatty acids											
C16:0	27.2	27.4	26.4	27.8	0.70	0.52	0.38	24.9	29.5	0.48	<0.01
C18:0	4.29	4.43	4.33	4.41	0.143	0.70	0.70	4.78	3.95	0.089	<0.01
C18:1n-9	35.3	36.1	35.8	34.8	0.83	0.51	0.38	37.2	33.8	0.57	<0.01
C18:2n-6	11.45	9.63	12.08	11.74	0.412	0.09	0.09	12.20	10.30	0.245	<0.01
C18:3n-6	0.14	0.12	0.15	0.13	0.012	0.93	0.87	0.20	0.08	0.008	<0.01
C18:3n-3	1.08	1.12	1.16	1.32	0.057	0.06	0.53	1.25	1.09	0.036	<0.01
C20:3n-3	0.11	0.11	0.11	0.09	0.010	0.76	0.48	0.11	0.10	0.006	0.06
C20:4n-6	0.52	0.50	0.55	0.55	0.022	0.16	0.53	0.65	0.41	0.014	<0.01
C20:5n-3	0.09	0.09	0.08	0.08	0.006	0.55	0.74	0.09	0.07	0.003	<0.01
C22:5n-3	0.23	0.22	0.21	0.22	0.018	0.77	0.61	0.26	0.18	0.010	<0.01
<i>n</i> -3 ⁴	1.75	1.59	1.50	1.48	0.082	0.19	0.60	1.72	1.44	0.051	<0.01
<i>n</i> -6 ⁵	12.1	10.3	12.8	12.4	0.43	0.09	0.10	13.0	10.8	0.26	<0.01
Saturated	36.1	36.4	35.3	37.1	0.76	0.57	0.40	33.8	38.6	0.52	<0.01
Mono-unsaturated	49.1	50.8	49.5	48.1	0.61	0.71	0.04	50.4	48.3	0.42	<0.01
Poly-unsaturated	14.8	12.8	15.2	14.9	0.53	0.22	0.14	15.7	13.1	0.32	<0.01

324 ¹T13 = Sow's lactation diet containing 13% of hemicelluloses; T11 = Sow's lactation diet containing 11% of hemicelluloses; T9 = Sow's lactation diet containing
325 9% of hemicelluloses; T8 = Sow's lactation diet containing 8% of hemicelluloses.

- 326 ²Contrasts: L = Linear; Q = Quadratic.
- 327 ³ Days: d3 = Day 3 of lactation; d17 = Day 17 of lactation;
- 328 ⁴ $n-3$: sum of C18:3 $n-3$, C20:3 $n-3$, C20:5 $n-3$, C22:5 $n-3$.
- 329 ⁵ $n-6$: sum of C18:2 $n-6$, C18:3 $n-6$ and C20:4 $n-6$.

330 ***Volatile fatty acid concentrations in milk***

331 The VFA concentration and the proportion of butyrate linearly increased ($P < 0.01$;
332 Table 6) with decreased HC content in the maternal diet, resulting in an increased in
333 total VFA by 25% and butyrate proportion by 60%. Regardless of the dietary treatment,
334 total VFA concentration decreased ($P \leq 0.05$) by 71% between days 3 and 17. The
335 proportion of methanoate increased ($P < 0.01$), and the proportion of acetate
336 decreased ($P < 0.01$) between days 3 and 17, whereas the levels of propionate,
337 isobutyrate, butyrate and isovalerate remained unchanged.

Table 6. Effect of decreasing hemicellulose levels in sow's lactation diet on the volatile fatty acid profile of milk

Item	¹ Dietary Treatments				SEM	² Contrasts		³ Stage of lactation		SEM	P-value
	T13	T11	T9	T8		L	Q	d3	d17		
Total volatile fatty acids, mmol/kg	3.07	3.58	3.60	3.86	0.28	0.03	0.60	4.12	2.94	0.19	<0.01
Proportion of individual VFA, %											
Methanoate	9.41	9.50	9.38	9.93	0.287	0.94	0.28	9.16	9.95	0.187	<0.01
Acetate	88.90	89.00	88.90	88.30	0.353	0.31	0.17	89.21	88.36	0.220	<0.01
Propionate	0.30	0.30	0.25	0.20	0.041	0.19	0.84	0.25	0.28	0.026	0.29
Isobutyrate	0.04	0.04	0.05	0.03	0.007	0.86	0.79	0.04	0.05	0.004	0.17
Butyrate	0.53	0.60	0.75	0.86	0.153	<0.01	0.64	0.68	0.69	0.104	0.29
Isovalerate	0.76	0.55	0.57	0.57	0.080	0.80	0.21	0.61	0.61	0.043	0.81

339 ¹T13 = Sow's lactation diet containing 13% of hemicelluloses; T11 = Sow's lactation diet containing 11% of hemicelluloses; T9 = Sow's lactation diet containing
340 9% of hemicelluloses; T8 = Sow's lactation diet containing 7% of hemicelluloses.

341 ²Contrasts: L = Linear; Q = Quadratic

342 ³d3 = Day 3 of lactation; d17 = Day 17 of lactation

343 **Discussion**

344 ***Effect of decreasing the level of hemicelluloses on sows' performance***

345 Excluding fibre intake, the sow's performances were not affected by dietary HC. In the
346 present study, one goal was to have a similar total DF intake among the sows in the
347 four treatments but different intakes of IDF and SDF. This objective was only partially
348 achieved as there was no linear effect but a quadratic effect for total DF intake.
349 Nonetheless, due to similar feed intake during the pre-farrowing and lactation periods,
350 decreasing the level of HC also reduced the intake of the low-molecular-weight DF and
351 SDF fractions. Similar to the present study, Shang et al. (2021) found no effect either
352 on sow's BW or backfat thickness at farrowing and weaning when the dietary SDF level
353 was decreased from 40.6 g/kg to 13.9 g/kg in the late gestation and from 27.2 g/kg to
354 14.3 g/kg during lactation. In addition, considerably high SDF intake can negatively
355 affect litter performance. Indeed, Liu et al. (2020) reported that from day 90 of gestation
356 to farrowing, a daily intake of 215 g of SDF (SDF: 45.7 g/kg as fed), compared with
357 138 g/day (29.7 g/kg as fed) and 96 g/day (17.8 g/kg), decreases the number of piglets
358 and litter weight at weaning. In the present study, sows received between 133 and 83
359 g/day of SDF according to the diets, from day 110 of gestation to farrowing. Therefore,
360 compared to the study of Liu et al. (2020), the SDF intake during this period for the four
361 treatment groups was not sufficiently elevated to negatively impact litter performances.

362 ***Effect of decreasing hemicelluloses levels on milk composition and milk VFA*** 363 ***profile***

364 Milk yield and composition play a crucial role in the growth of suckling piglets to reach
365 an adequate weaning weight. In the present study, decreasing the level of HC in the

366 maternal diet affected milk composition but did not affect milk yield. Furthermore,
367 lactose content decreased, whereas copper and threonine proportions increased with
368 decreased HC level. A previous study showed that glucose, glycerol and other glucose
369 precursors play an important role in the synthesis of lactose in sow's milk (Boyd et al.,
370 1995). Houdijk et al. (2002) reported that the fermentation of SDF occurs already at
371 the end of the ileum. As decreasing the level of HC also decreased the intake of SDF,
372 one can hypothesize that lowering the HC supply reduced the absorbed HC
373 fermentation products available for lactose synthesis. Moreover, due to the osmotic
374 power of lactose (Costa et al., 2019), milk yield may drop together with lactose as the
375 HC level decreases. Surprisingly, milk yield only decreased numerically, and this result
376 could be due to the differences in lactose concentration between the experimental
377 groups, which were not sufficiently large to affect milk yield. A further interest in the
378 present study is the linear increase in copper in milk with a decreased HC level. Copper
379 is an essential microelement for animals, with many biological functions, including iron
380 metabolism, immunity, protection from oxidative stress and improvement in the activity
381 of digestive enzymes (Huang et al., 2015). The milk concentration of copper is affected
382 by the source of the micromineral (Peters et al., 2010). However, as the same
383 micromineral source was used among the four dietary treatments, the mechanism
384 underlying the increase in copper concentration remains unclear. Similarly, with
385 decreased HC levels in the diet, the proportion of threonine in the milk increased. This
386 effect remains unclear, as the calculated digestible threonine levels were similar
387 between the T13 and T8 diets. Besides a similar DF content, hypothetically, decreasing
388 the HC level using several DF sources may affect the fermentation patterns in the gut,
389 namely, the concentration and proportion of VFA. As VFA can be absorbed,
390 transported through the blood and finally reach the mammary glands, modifications in

391 the milk composition are expected (Tian et al., 2020). Decrease in HC level increased
392 total VFA concentration and butyrate proportion in milk. Zhao et al. (2019) showed a
393 positive correlation between VFA concentration in pig's ileum and decreased HC level.
394 Given that sows can ferment DFs better than growing pigs, a similar phenomenon may
395 have occurred in the ileum of sows fed with a low HC level (Noblet and Le Goff, 2001).
396 Furthermore, this effect on VFA in milk may also be due to differences in the intake of
397 other DF fractions. As previously mentioned, decreasing HC level concomitantly
398 increased ADF intake and decreased SDF intake. A positive correlation was reported
399 between the ADF level in pig's diet and butyrate concentration in the faeces (Zhao et
400 al., 2019). In the present study, hypothetically, increased ADF intake in sows fed with
401 decreasing level of HC might have increased the butyrate proportion in the faeces and
402 then in the milk. Compared with IDF, SDF is rapidly fermented by bacteria, thereby
403 enhancing the production of VFA (Jha and Berrocso, 2015). Therefore, with
404 decreased SDF intake, VFA production should be lowered. However, the present study
405 showed that this concept was not evident and confirmed the importance of the source
406 of DF, as reported by some authors (Theil et al., 2014). Therefore, to understand the
407 effects of DF on milk composition, different fractions of DF, including HC and ADF
408 contents, must be considered.

409 ***Effects of the lactation diet on piglets' performance***

410 In the present study, modifying the level of HC in the maternal diet did not enhance
411 litter performance. This result is consistent with the results of Loisel et al. (2013), which
412 showed that modifying the maternal diet is easier to positively affect the performances
413 of L-BtW piglets than the performance of the litter overall. Therefore, decreasing the
414 HC level improved post-weaning performance and reduced the occurrence of

415 diarrhoea in the L-BtW piglets. By contrast, why the performance of N-BtW was
416 unaffected by the HC level even though the occurrence of diarrhoea increased in this
417 group remains unclear. The L-BtW piglets usually exhibit poor performances, such as
418 a high mortality rate and low ADG, which represents high economic costs for farmers
419 due to reduced slaughter weight and increased occupancy of the stables (López-Vergé
420 et al., 2018). Girard et al. (2021) highlighted the importance of early-life interventions
421 to improve the post-weaning development and health of this sub-population of piglets.
422 In the present study, the beneficial effects observed in L-BtW piglets during post-
423 weaning period like the improved growth performance and the lower incidence of
424 diarrhoea may be related to the combination of an increased relative abundance of
425 butyrate, threonine and copper and to an increased concentration of total VFA in milk.
426 Given that piglets are highly susceptible to intestinal bacterial disorders during the
427 post-weaning period, butyrate, due to its recognised role in gut health, could have been
428 useful in increasing gut impermeability, alleviating diarrhoea in L-BtW piglets during
429 the second week post-weaning (Feng et al., 2018). In addition, increasing threonine
430 and copper proportions in the milk in the pre-weaning period can help accelerate the
431 gut maturation of those piglets (Lalles et al., 2009). Threonine plays a critical role in
432 the regulation of intestinal mucosal integrity, as it is required for the production of
433 mucins and immunoglobulins, improving the physical protection from the attachment
434 of microbes to the mucosal surface (Van Klinken et al., 1995). By contrast, copper can
435 help against pathogenic bacteria because of its bacteriostatic properties, which affect
436 the community structure of microorganisms in the caecum and colon (Højberg et al.,
437 2005). A lower relative abundance of *Alistipes*, *Lachnospiraceae*, *Ruminococcaceae*
438 and *Prevotellaceae* has been reported in the colon and ileum of L-BtW piglets
439 compared with N-BtW piglets (Li et al., 2019). These genera enhance gut health and

440 immune functions in the host (Den Besten et al., 2013). Given that, colostrum and
441 mature milk are key components in shaping piglet microbiota (Trevisi et al., 2021), the
442 modification of milk composition induced by decreased HC level in the sow diet might
443 have changed the gut microbiota of L-BtW piglets and improved their health and
444 growth.

445 ***Effect of lactation stage on milk composition***

446 Sow's milk composition is strongly affected by changes throughout the lactation period.
447 Transitional milk (48–72 h after parturition) contain higher amounts of lipids, protein
448 and dry matter compared with mature milk (from day 10 of lactation) (Csapó et al.,
449 1996). In the present study, the passage from transitional milk to mature milk was
450 characterised by a decrease in protein and ash contents and an increase in lactose
451 content. Nevertheless, the contents of fat, dry matter and gross energy decreased only
452 numerically from day 3 to day 17. Indeed, in the present experiment, the lack of
453 statistical differences on those traits is in disagreement with several studies (Csapó et
454 al., 1996; Theil et al., 2014), where differences between the sampling days were
455 reported. This might be related to differences in sow genotypes, sow management and
456 litter size between the present study and the previous ones. Similarly, the decrease in
457 amino acid proportion follows the same trend as protein content, except for glycine,
458 lysine and methionine, which remained stable over lactation, and for glutamate and
459 proline, which increased from day 3 to day 17. Therefore, the high level of amino acids
460 in transitional milk reflects the protein level, mainly because of the high content of
461 immunoglobulins (Klobasa et al., 1987). The mineral content was also affected by the
462 stage of lactation with an increase in the calcium level and a decrease in the potassium
463 and zinc levels from transitional milk to mature milk in agreement with Csapó et al.

464 (1996). Moreover, the phosphorus content decreased between days 3 and 17. The
465 reason for this decrease over lactation remains unclear but might be related to a
466 dilution effect, as it follows the numerical decrease in dry matter. When expressed per
467 kilogram of dry matter, the phosphorus concentration was similar between days 3 and
468 17. Moreover, from transitional milk to mature milk, the decrease in the proportion of
469 mono- and polyunsaturated fatty acids and the increase in the proportion of saturated
470 fatty acids are related to changes in the proportion of individual fatty acids. The
471 increase in C16:0 proportion and decrease in the proportions of C18:0, C18:1n-9,
472 C18:2n-6, C18:3n-6, C18:3n-3, C20:4n-6 and C20:5n-3 observed in the present study
473 have already been described in a previous study (Hu et al., 2019). Furthermore, Hu et
474 al. (2019) reported a positive correlation between calcium and C16:0 fatty acid.

475 In conclusion, when the DF level is the same, feeding lactating sows with a lower HC
476 level can positively affect the milk composition and the development of L-BtW piglets.
477 As HC content decreased, the growth performance of the L-BtW piglets improved after
478 weaning, and the occurrence of diarrhoea decreased, particularly in the second week
479 post-weaning. Moreover, it increased the proportion of butyrate, copper and threonine
480 and increased the VFA concentration in the milk. Therefore, this study highlighted the
481 importance of the maternal diet in lactation to positively affect the development and
482 health of L-BtW piglets in the post-weaning period.

Ethics approval

The experiment was conducted in accordance with the Swiss Guidelines for Animal Welfare, and the Swiss Cantonal Committee for Animal Care and Use approved all procedures involving animals (approval number: 2019_25_FR).

Data accessibility

The data that support the findings of this study are publicly available in Zenodo (<https://doi.org/10.5281/zenodo.5814624>).

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Author contributions

Francesco Palumbo and Marion Girard validated the data and carried out the main statistical analyses. Marion Girard and Giuseppe Bee conceived the study design and secured substantial funding. Francesco Palumbo and Marion Girard performed the animal experiment, recorded the data and collected and processed the milk samples. Marion Girard, Francesco Palumbo, Giuseppe Bee and Paolo Trevisi supervised analyses and drafted and critically reviewed the manuscript. All authors read and approved the final manuscript.

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Declaration of interest

The authors declare they have no conflict of interest relating to the content of this article.

Supplementary materials

The Supplementary Table S1 and the statistical codes used can be found in the Supplementary Materials.

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