1	Decreasing the level of hemicelluloses in sow's lactation diet affects the milk
2	composition and post-weaning performance of low birthweight piglets.
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10 Abstract

Hemicelluloses (HC) are polysaccharides constituents of the cell walls of plants. They 11 are fermented in the gut to produce volatile fatty acids (VFA). The present study 12 investigated the effects of decreasing HC level in sow's lactation diet on sow 13 performances, offspring development and milk composition. From 110 days (d) of 14 gestation until weaning (26±0.4 d post-farrowing), 40 Swiss Large White sows were 15 assigned to one of the four dietary treatments: (1) T12 T13 (HC: 121 127g/kg), (2) T11 16 (HC: 107.6 114g/kg), (3) T9 (HC: 86.4 94g/kg) and (4) T7 T8 (HC: 71.9 80g/kg). Milk 17 was collected at 3 and 17d of lactation. At birth, piglets were divided into two groups 18 according to their birthweight (BtW): normal (N-BtW; BtW > 1.20 kg) or low (L-BtW; 19 BtW \leq 1.20 kg). Decreased HC levels in the maternal diet linearly increased ($P \leq 0.05$) 20 the body weight of L-BtW piglets at two weeks post-weaning and linearly decreased (P 21 \leq 0.05) diarrhoea incidence and duration in this category. The concentrations of 22 copper, threonine and VFA, as well as the proportion of butyrate, in milk linearly 23

increased ($P \le 0.05$), whereas lactose content linearly decreased ($P \le 0.05$) with decreased HC in the maternal diet. The present study provides evidence that decreasing HC level in sow's lactation diet can positively affect the composition and VFA profile of milk and ultimately favour the growth and health of L-BtW piglets.

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29 Keywords: Dietary fibres, lactose, pigs, volatile fatty acids, butyrate

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31 Introduction

Hemicelluloses (HC) represent a complex group of polysaccharides present in the cell 32 walls of all plants, consisting mainly of pentoses (D-xylose and D-arabinose), hexoses 33 (D-galactose, D-glucose and D-mannose) and uronic acids that can be estimated as 34 the difference between NDF and ADF (Huang et al., 2021; Van Soest et al., 1991). As 35 part of dietary fibres (DF), they can resist digestion by endogenous enzymes of the 36 gut. Thus, they can reach the large intestine and promote the growth and activity of 37 beneficial bacteria that produce volatile fatty acids (VFA) (Lattimer and Haub, 2010). 38 These latter, namely, acetate, propionate and butyrate, provide up to 28% of the 39 energy requirements in growing pigs and even more in sows, where they can be 40 absorbed and transferred to the milk and serve as an energy source for milk synthesis 41 (Noblet and Le Goff, 2001; Tian et al., 2020). A previous study focused on increasing 42 the level of DF in sow's gestation diet showed that adding up to 20% DF increases 43 colostrum fat content, as well as colostrum intake, of low birthweight (BtW) piglets (L-44 BtW) ($0.6 \text{ kg} \ge \text{BtW} < 0.9 \text{ kg}$) and decreases litter mortality during the suckling period 45 (Loisel et al., 2013). To our knowledge, the effects of HC level have been investigated 46 in growing pigs but never in lactating sows (Zhao et al., 2019).on lactating sows have 47

never been investigated However, this aspect can be relevant, since sow nutrition 48 inclusion of DF during lactation may also positively affect the development and gut 49 health of piglets (Paßlack et al., 2015). Like all DF, HC can be differentiated according 50 to their physiochemical properties such as their solubility in water and their intestinal 51 fermentability. Indeed, depending on the plants, HC might be considered as a source 52 of soluble dietary fibres (SDF) (Jiménez-Escrig et al., 2000). Compared to insoluble 53 dietary fibres (IDF), SDF are easily fermented and may be completely degraded in the 54 end of small intestine (Houdijk et al., 2002). Due to their slower fermentability, the 55 majority of the IDF reach the large intestine, which acts as a fermentation chamber 56 producing VFA (Lattimer and Haub, 2010). Therefore, the present study aims to fill this 57 gap by comparing four diets characterised by similar total DF and crude fibre contents 58 but different HC levels by varying the sources of DF. We hypothesised that decreasing 59 the level of HC in sow's lactation diet may affect gut fermentation particularly in the 60 large intestine and ultimately modify milk composition. 61

62 Materials and methods

63 Animals, housing and treatments

The experiment was conducted during late gestation and lactation of 40 Swiss Large 64 White sows from five farrowing batches. Approximately 10 days before the expected 65 time of farrowing, sows were moved to farrowing rooms arranged with individual 7.1 66 m² farrowing crates, consisting of a 5.89 m² concrete solid floor and a 1.21 m² concrete 67 slatted floor. Each crate was equipped with an electronic sow feeder (Schauer Spotmix, 68 Schauer Agrotronic GmbH, Austria), nipple drinker and a heated covered area for 69 piglets. The ambient temperature was maintained at 24 °C, and artificial lights were on 70 from 0800 h to 1700 h. On day 110 of gestation, the sows were randomly allocated to 71

one of the four experimental lactation diets based on parity (mean \pm SEM: 3.5 \pm 0.7) 72 and BW (mean ± SEM: 286.5 ± 13.6 kg). Parturition was induced when gestation period 73 exceeded 116 days with an intramuscular injection of 1 ml (0.25 g/ml) of cloprostenol 74 (Estrumate®, MSD Animal Health GmbH, Luzern, Switzerland). Within the first 24 h 75 following birth, piglets were identified by an individual ear tag and received iron 76 injection (Feridex® 10%, AMAG Pharmaceuticals, Inc., Waltham, USA). Piglets 77 weighing less than 800 g at birth were excluded from the experiment. To adjust litter 78 size to an average of 12 piglets per sow, cross-fostering was carried out only on male 79 piglets 24 h post-farrowing. After anaesthetisation, the male piglets were castrated in 80 81 the second week. Piglets were weaned on day 25.7 ± 0.44 (mean \pm SEM) of age but were kept in their respective farrowing crates until 2 weeks post-weaning. The heating 82 nest temperature was set at 40 °C following birth and then gradually decreased by 0.5 83 °C per day to reach a final temperature of 32 °C 84

85 **Diets and feeding**

The experimental diets were formulated to be isonitrogenous and isocaloric (Table 1) 86 and to differ in DF sources and HC content: (1) T12 T13 (HC: 121 127 g/kg), (2) T11 87 (HC: 107.6 114g/kg), (3) T9 (HC: 86.4 94g/kg) and (4) T7 T8 (HC: 71.9 80g/kg). The 88 daily feed allowance was calculated according to the current Swiss feeding 89 recommendations for pigs (Agroscope, 2018). Sows had ad libitum access to water 90 and were provided with moderate quantities of straw bedding, as required by the Swiss 91 legislation. During the end of gestation, feed allowance was on average 3.04 ± 0.16 kg 92 $(mean \pm SEM)$. While, during lactation, the feed allowance was gradually increased by 93 0.5 kg/day until ad libitum feeding on day 12 of lactation approximatively. All diets were 94 delivered three times per day in three equal meals using a computerised feed delivery 95

96 system (Schauer Spotmix, Schauer Agrotronic GmbH, Austria). Throughout the 97 experiment, the feed refusals of the sows were weighed daily to calculate actual feed 98 intake. From day 18.7 ± 0.44 of age (mean \pm SEM) to 2 weeks post-weaning (mean \pm 99 SEM: day 39.7 ± 0.44 of age), piglets had *ad libitum* access to a post-weaning standard 100 starter diet and water. The post-weaning starter diet contained 170 g/kg crude protein, 101 58 g/kg fat, 50 g/kg crude fibre and 14 MJ/kg digestible energy.

		Dietary Trea	atments ¹	
Item	T12 -T13	T11	Т9	T7- T8
Ingredients (%)				
Barley, ground	54.4	38.7		4.7
Oat flakes			4.0	18.2
Corn, ground	10.3		26.9	16.0
Rye		25.0	10.0	
Wheat, ground			13.1	15.0
Wheat starch	4.0	4.0	4.0	4.0
Molasses				4.0
Animal fat RS 65	2.4	2.4	3.0	3.8
Potato protein	10.0	10.0	10.0	10.0
Soybean meal	10.0	10.0	10.0	10.0
Flaxseed Meal	0.6			
Rapeseed meal		0.4		1.7
Oat hulls			4.0	8.0
Lupin			2.5	
Wheat bran			4.0	
Beet pulp	3.0	5.0	4.0	
L-lysine-HCL	0.070	0.057	0.057	0.056
DL-methionine	0.200			
L-threonine	0.050			0.050
L-tryptophan	0.020	0.006	0.013	0.003
Dicalcium phosphate	0.94	0.70	0.82	0.85
Calcium carbonate	1.57	1.38	1.39	1.47
Salt	0.59	0.52	0.42	0.41
Pellan ²	0.40	0.40	0.40	0.40
Celite	1.00	1.00	1.00	1.00

Table 1. Ingredients and composition of the sow's lactation diet

Premix ³	0.40	0.40	0.40	0.40
Natuphos 5000 G ⁴	0.01	0.01	0.01	0.01
Gross chemical composition analysed (g/kg as fed)				
Dry matter	900	894	897	900
Crude protein	193	191	192	196
Fat	51	46	57	60
Crude fibre	43	43	47	46
Ash	63	61	60	63
NDF	184	174	163	154
ADF	63 57	67 60	76 69	82 79
Hemicelluloses ⁵	121 -127	108- 114	86 94	72- 80
Total dietary fibres	210	227	220	203
Low-molecular-weight dietary fibres	18	23	18	14
Soluble dietary fibres	43	44	35	28
Insoluble dietary fibres	149	160	167	161
IDF/SDF ⁶	3.46	3.63	4.77	5.75
Calcium	9.4	9.4	9.3	8.7
Phosphorus	5.0	4.6	5.0	4.7
Gross chemical composition calculated				
Digestible energy (MJ/kg)	14.1	14.1	14.1	14.1
Digestible phosphorus (g/kg as fed)	3.1	2.8	2.8	2.8
Digestible essential amino acids (g/kg as fed)				
Lysine	9.6	9.6	9.6	9.6
Methionine	4.9	2.9	3.0	3.0
Threonine	6.9	6.3	6.4	6.9
Tryptophan	2.0	1.8	1.8	1.8

103 1712 T13= Sow's lactation diet containing 42 13% of hemicelluloses; T11 = Sow's lactation diet

104 containing 11% of hemicelluloses; T9 = Sow's lactation diet containing 9% of hemicelluloses; T7 T8 =

105 Sow's lactation diet containing 78% of hemicelluloses.

²Pellet binding aid: Pellan, Mikro-Technik, Bürgstadt, Germany.

³Supplied per kg of diet: vitamin A, 8000 IU; vitamin D3, 800 IU; vitamin E, 40 mg; menadione, 2 mg;

thiamine, 2 mg; riboflavin, 5 mg; biotin, 0.1 mg; niacin, 20 mg; pantothenic acid, 20 mg; iodine (as

109 calcium iodate), 0.55 mg; copper (as copper sulphate), 7 mg; manganese (as manganese oxide), 20

110 mg; zinc (as zinc oxide), 55 mg; selenium (as sodium selenite), 0.2 mg.

⁴Phytase supplemented with 500 units of *Aspergillus niger* phytase/kg diet.

⁵Hemicellulose: calculated as the difference between NDF and ADF.

⁶Ratio of insoluble to soluble dietary fibres

114 Sow and piglet performance

The BW of the sows, body condition score (BCS) and backfat thickness were recorded 115 at the 110th day of gestation and during on the day of farrowing and weaning. Weight 116 loss during lactation was calculated as the weight difference between farrowing and 117 weaning. Based on visual observation and palpations, BCS was determined according 118 to a scale ranging from 1 (very thin) to 6 (obese) points (Dourmad et al., 2001), 119 including intermediate values of 0.33 points. Briefly, the trained personnel assessed 120 sows by palpating the shoulders, ribs, backbone and hips, followed by a visual 121 observation. Backfat thickness was measured on each side at 65 mm of the dorsal 122 midline at the level of the last rib (P2) using a digital ultrasound back-fat indicator 123 124 (Renco Lean Meter Digital Backfat Indicator, Renco Corporation, Minneapolis, Minnesota, USA). Backfat thickness loss during lactation was then calculated as the 125 difference between backfat thickness measurements during farrowing and weaning. At 126 farrowing, the number of born alive, stillborn and mummified piglets were recorded 127 within each litter. Farrowing was recorded using a digital video recorder to estimate the 128 farrowing duration, which is defined as the time span between the time of birth of the 129 first and last piglet of the litter. Furthermore, the time between the start of the farrowing 130 and the first piglet suckling was also recorded. At birth, the piglets were individually 131 weighed, and crown-to-rump length and body circumference were recorded. Piglets 132 were then individually weighed 5 and 16 days postpartum, during weaning (mean ± 133 SEM: 25.7 ± 0.44 days of age) and at 1 (mean \pm SEM: 32.7 ± 0.44 days of age) and 2 134 weeks post-weaning (mean ± SEM: 39.7 ± 0.44 days of age). The average daily gain 135 (ADG) and litter weight during birth and weaning were calculated from these data. Milk 136 yield was calculated as the individual piglet gain summed in the same litter multiplied 137 by a numerical coefficient of 4.2 (Van der Peet-Schwering et al., 1998). The indices of 138 body conformation were calculated based on the measurements of the individual 139

birthweight (BtW) and the crown-to-rump length. The body mass index was calculated 140 as the ratio of BtW to the squared value of the crown-to-rump length, and the ponderal 141 index was calculated as the ratio of BtW to the cubic value of the crown-to-rump length 142 (Hales et al., 2013). In addition, piglets were divided into two BtW groups: normal (N-143 BtW; BtW > 1.20 kg) or low (L-BtW; BtW \leq 1.20 kg). From 1 week before weaning 144 onwards, feed intake and refusals (including feed waste) per pen as well as the 145 occurrence of diarrhoea were recorded daily. Diarrhoea incidence percentage was 146 determined according to a daily faecal score assessed using a scale from 0 = no147 *diarrhoea* to 1 = *diarrhoea*. The percentage of diarrhoea per group was calculated as 148 the sum of piglets with a faecal score of one divided by the total number of piglets. 149

150 Sample Collection

151 Within each farrowing series, feed samples of the four diets were collected weekly and pooled over the experimental period to determine the chemical composition. On days 152 3 and 17 of lactation, milk samples were manually collected from all functional teats 153 after an intramuscular injection of 2 ml of oxytocin (Intertocine-S, MSD Animal Health 154 GmbH, Luzern, Switzerland). Before milking, the piglets were temporarily isolated from 155 the sow for 2 h, and the teats were cleaned with humid wipes. One aliguot of milk was 156 refrigerated at 5 °C with 4 mg of bronopol to determine somatic cell concentration, and 157 three aliquots were immediately stored at -20 °C for further analysis. 158

159 Analytical Methods

160 Feed Analysis

After being ground to pass a 1-mm screen (Brabender rotary mill; Brabender GmbH & Co. KG, Duisburg, Germany), feed samples were analysed for dry matter content by heating at 105°C for 3h followed by incineration at 550°C until a stable mass was

reached to determine the ash content according to ISO 5984:2002 (prepASH, Precisa 164 Gravimetrics AG, Dietikon, Switzerland). An inductively coupled plasma optical 165 emission spectrometer (ICP-OES, Optima 7300 DV; Perkin-Elmer, Schwerzenbach, 166 Switzerland) was used to measure mineral content (European Standard EN 167 15510:2008). The CP content was calculated as nitrogen (N) content multiplied by a 168 coefficient of 6.25, where N was determined with the Dumas method (ISO 16634-169 1:2008). Fat content was extracted with petrol ether after acid hydrolysis (ISO 170 6492:1999). Different categories of fibres were analysed by standard protocols. Crude 171 fibre content was determined gravimetrically (ISO 6865:2000) by incineration of 172 residual ash after acid and alkaline digestions using a fibre analyser (Fibretherm 173 Gerhardt FT-12, C. Gerhardt GmbH & Co. KG, Königswinter, Germany). The NDF and 174 ADF contents (ISO 16472:2006 for NDF and ISO 13906:2008 for ADF) were analysed 175 with the same fibre analyser (Fibretherm Gerhard FT-12, C. Gerhardt GmbH & Co. 176 KG, Konigswinter, Germany) and were expressed without residual ash. NDF 177 determination was evaluated with heat stable amylase and sodium sulfite and 178 expressed without residual ash after incineration at 600°C for 3 h. Soluble (The 179 contents of-SDF), insoluble (IDF) and low-molecular-weight DF contents were 180 measured according to AOAC Method 2011.25, and the total DF content was 181 calculated as the sum of the three aforementioned types of DFs. 182

183 Milk Analysis

The dry matter of the frozen milk samples was determined after freeze-drying (Christ DELTA 2-24 LSC, Kühner AG, Birsfelden, Switzerland) for 70 hours. Subsequently, freeze-dried samples were milled with a mortar. Residual dry matter, ashes, mineral and CP nitrogen contents were analysed as previously described for the feed chemical

analysis, except that a multiplicative coefficient of that CP was expressed as N x 6.38 188 was applied for the calculation of CP. Except for tryptophan, all amino acids were 189 determined as described in ISO 13903:2005. Briefly, after oxidation, 24 h of acid 190 hydrolysis occurred with 6M HCI and derivatization with AccQ-Tag Ultra reagent 191 (Waters corporation, Milford, USA USA), the amino acid profile was determined by 192 ultra-high-performance liquid chromatography (UHPLC) coupled with a UV detector 193 (Vanguish, Thermo Scientific, Reinach, Switzerland. Tryptophan content was 194 quantified by HPLC (LC 1290 Infinity II LC System, Agilent Technologies, USA) 195 according to ISO 13904:2016. Gross energy content was determinate by combustion 196 197 in a calorimetric vessel under pure oxygen condition using an adiabatic bomb calorimeter (AC600 Semi-Automatic Calorimeter, Leco Corporation, USA) (ISO 198 9831:1998). Lactose content was determined by enzymatic testing with β -199 galactosidase and galactose dehydrogenase (Enzytec TM Liquid Lactose/D-Galactose 200 Ref. No. E8110, R-Biopharm AG, Darmstadt, Germany). Somatic cells count (ISO 201 13366-2) was determined by flow cytometry (Somacount FC, Bentley Instruments Inc., 202 USA). Fatty acid methyl esters, as described by Kragten et al. (2014), and the VFA 203 profile (ISO 15884:2002) (ISO 15885:2002) were determined by gas-liquid 204 chromatography (Gaschromatograph Series II Agilent 6850, Agilent Technologies 205 2000, USA and Gaschromatograph Serie Agilent 6890, Agilent Technologies 2000, 206 USA, respectively). Fat content was determined as total fatty acids multiplied by a 207 coefficient of 1.05. 208

209 Statistical Analysis

Due to health problems that could not be related to the dietary treatment, one T9 sow was excluded from the experiment. Data were analysed by ANOVA using the 'lme' and

the 'glmmPQL' function of the *nlme* package of R Studio (version 4.0.2 for Windows). 212 Regarding sow performance, milk composition and VFA profile, the sow was the 213 experimental unit; the pen was the experimental unit regarding piglet feed intake and 214 litter performance; and the piglet was the experimental unit of piglet's individual 215 performance, days and percentage of diarrhoea. Linear regression models, including 216 the treatment and the farrowing batch as fixed effects, were used to fit data related to 217 sow performance, litter performance, piglet feed intake and days with diarrhoea. Data 218 related to piglets' individual performance were analysed using a linear mixed-effects 219 model, including the treatment and the farrowing batch as fixed effects and the sow as 220 221 random effects. Milk composition and VFA profile were analysed with a linear mixedeffects model and fitted in repeated measurements, including the treatment, the 222 farrowing batch, the sampling day, and the interaction between the treatment and 223 sampling day as fixed effects and the sow as random effect. Before analysis, 224 logarithmic transformation was applied to the milk fatty acid and milk VFA data due to 225 the non-normality of the residuals. The percentage of diarrhoea was analysed using a 226 generalised linear mixed model using Penalized Quasi-Likelihood, including the 227 treatment, the farrowing batch and the day as fixed effects and the piglet as a random 228 229 factor. Orthogonal polynomial contrasts were implemented to evaluate the linear or quadratic effects of decreasing HC level. The results are expressed as the least square 230 means \pm SEM. Linear and quadratic effects were considered significant at $P \le 0.05$. 231

232 Results

233 Sows' performance

The sow BW, BCS and backfat thickness on day 110 of gestation and during farrowing 234 and weaning were not influenced by the dietary treatment, resulting in similar weight 235 and backfat thickness losses during the lactation period. Daily feed intake in the pre-236 farrowing period and during lactation did not differ between treatments. Fibre intake 237 was partially influenced by dietary treatments. In both the pre-farrowing and lactation 238 periods, the NDF, HC, (linear effects; P < 0.01), low-molecular-weight DF and SDF 239 intake decreased (linear and quadratic effects; P < 0.01), and the ADF intake increased 240 (linear effect; P < 0.01) with decreasing HC levels in the diet. A quadratic effect ($P \le$ 241 0.04) of the HC level was found in the diets on the intake of total DFs in the pre-242 farrowing and lactation periods. At birth, litter traits, such as total born, born alive and 243 stillborn piglets, did not differ, leading to comparable litter weights in the four 244 treatments. Likewise, the dietary treatments had no effect on the total number of piglets 245 weaned and, consequently, on litter weight at weaning. Farrowing duration and the 246 time between the start of the farrowing and the first piglet suckling were was not 247 influenced by dietary treatments. During the entire lactation period, milk yield was not 248 influenced by the dietary treatments, with an average estimated production of 10.38 249 kg/day per sow (Table 2). 250

Table 2. Effect of decreasing hemicelluloses level in lactation diet on sow's
 performance

			² Contrasts				
Item	T12 T13	T11	Т9	17 -T8	SEM	L	Q
Sows							
Number of sows, <i>n</i>	10	10	9	10			
Range of parity, <i>n</i>	3.8	3.8	3.5	3.5	0.69	0.51	0.99
Farrowing duration, min	308	337	321	262	70.7	0.54	0.44
Body weight, kg							
D110	284	291	284	287	13.6	0.71	0.89
Farrowing	264	267	269	272	14.1	0.67	0.99
Weaning	233	238	248	246	12.5	0.39	0.78

Weight loss in lactation, kg BCS. <i>n</i>	30.6	28.7	20.9	26.1	2.58	0.19	0.30
D110	4.09	4.10	4.03	3.83	0.129	0.79	0.18
Farrowing	3.58	3.59	3.40	3.64	0.148	0.96	0.40
Weaning	2.71	2.62	2.81	2.94	0.246	0.42	0.62
Backfat thickness, mm					00	•••=	0.0-
D110		<u>15 8</u>	<u>14 8</u>	<u>12 7</u>			
2	13.8	14.8	12 7	15.8	0.88	0.31	0.23
Farrowing	13 7	14.6	12 7	15.6	0.88	0.34	0 24
Weaning	11.3	11.9	11.5	12.9	0.71	0.18	0.52
Backfat thickness loss in lactation mm	2.38	2.66	1 25	2 67	0.505	0.82	0.24
Milk vield kg/dav	10.61	10.85	10.09	9.97	0 720	0.02	0.79
Feed intake kg/day	10.01	10.00	10.00	0.07	0.720	0.11	0.70
Pre-farrowing	2 93	3 03	3 03	3 00	0 155	0 75	0.68
Lactation	5.67	5.00	5 77	5.87	0.100	0.70	0.00
Fibre intake a/day	0.07	0.00	0.11	0.07	0.201	0.00	0.70
Pre-farrowing							
Crude fibre	127	120	141	130	70	0 13	0 74
NDE	538	527	/02	161	26.1	0.13	0.74
	185	202	-32 230	2/18	11 3	0.05	0.03
ADI	160	192	200	240	10.2	<0.01	0.00
Hemicelluloses	353	325	200	22 4 013	15.1		0.90
Terricelidioses	270	345	201	210	16.1	<0.01	0.48
Total dictany fibras	614	699	667	610	22.1	0 82	0.04
Low molecular weight dictory fibro	52 52	70	55	10	00.1 00	0.02	0.04
Soluble dictory fibres	122	10	106	4Z 02	2.0 5.0	<0.01	<0.01
Soluble dietary fibres	100	120	506	00	0.0	0.01	0.01
	430	400	500	404	24.0	0.14	0.14
	246	054	260	070	10.4	0.06	0.01
	240	204	209	212	10.4	0.00	0.01
	1043	1033	937	903	41.3	<0.01	0.75
ADF	308 205	390 256	4 39 206	4 04 420	10.4 11 0	<0.01	0.71
	323	300	390	438	14.0		0.71
Hemicelluloses	000 740	030 677	4 90 544	4-10 465	20.0	<0.01	0.49
Total distant fibros	1100	1250	04 I 1070	400	20.0	0.76	0.02
l olai dielary libres	1190	100	1270	1190	01.1 4.6	0.70	0.02
Low-molecular-weight dietary libre	102	137	104	0Z 160	4.0	< 0.01	< 0.01
Soluble dietary fibres	244	201	202	103	9.0	<0.01	<0.01 0.00
Insoluble dietary libres	840	950	964	947	37.1	0.06	0.09
Sucking piglets	25	00	20	20	11.0	0.00	0.00
First pigiet suckling, min	30	Zð	20	38	11.0	0.98	0.23
Number of piglets per litter, n	40 5	40.7	40.5	44.0	4.40	0.05	0.70
	13.5	13.7	13.5	14.3	1.12	0.65	0.76
Born allve ³	12.8	12.4	11.4	12.7	1.27	0.82	0.49
Suiidorn	0.7	1.3	2.1	1.6	0.65	0.22	0.40
After cross-tostering	11.4	11.4	11.5	11.6	0.79	0.85	0.91
vveaned	10.7	10.9	11.3	10.7	0.76	0.94	0.60
Litter weight, kg							

253	¹ T12 T13 :	= Sow's la	ctation diet	containing 42	13% of he	emicellulose	s; T11 =	Sow's la	ctation di	ət
At	weaning			81.9	83.9	78.0	79.4	5.50	0.59	0.96
At	birth			20.5	20.5	21.1	20.3	1.63	0.99	0.81

containing 11% of hemicelluloses; T9 = Sow's lactation diet containing 9% of hemicelluloses; T7 T8 =

255 Sow's lactation diet containing 7.8% of hemicelluloses.

256 ²Contrasts: L = Linear; Q = Quadratic.

³ including piglets weighing less than 800g at birth.

258 Piglets' individual performance

Body characteristics, such as body circumference, crown-to-rump length, body mass 259 index and ponderal index, were not affected by the lactation diet of the sows (Table 3). 260 Similarly, piglet BW development, ADG and feed intake were not affected by the dietary 261 treatments. During the first week post-weaning, the incidence of diarrhoea and the 262 number of days with diarrhoea were similar among the treatments. By contrast, during 263 the second week of post-weaning, a quadratic increase ($P \le 0.05$) in the incidence of 264 diarrhoea and the number of days with diarrhoea was observed with decreasing HC 265 level. When focusing on the two BtW categories, the effect of the sow diets in the L-266 BtW group showed interesting observations (Table 4). Excluding the incidence of 267 diarrhoea and the number of days with diarrhoea that linearly increased during the 268 second week post-weaning (P < 0.01) with decreased HC level, no dietary effects were 269 observed in N-BtW piglets (Supplementary Table 1). The BtW, the BWs until one week 270 post-weaning and in accordance the ADG in this period were similar among the 271 experimental treatments for L-BtW piglets. By contrast, the decrease in HC level in the 272 sow diets increased (linear effect; $P \le 0.04$) the BW and the ADG in the second week 273 post-weaning and the overall ADG from birth to two weeks post-weaning of L-BtW 274 piglets. Likewise, the ADG of the L-BtW pigs remained similar among the dietary 275 treatments in the suckling period. Nevertheless, decreasing the HC level in the 276 maternal diet improved ($P \le 0.05$) the ADG of L-BtW piglets in the second week post-277

weaning and thereby the overall ADG of the L-BtW piglets during the experiment. In 278 the first week post-weaning, the dietary treatments did not affect either the incidence 279 of diarrhoea or the days with diarrhoea of L-BtW piglets. In the second week post-280 weaning, the incidence of diarrhoea and days with diarrhoea linearly decreased (P < 281 0.01) with decreased HC level in the maternal diet. Except for the linear increase in the 282 incidence of diarrhoea and increase in the number of days with diarrhoea in the second 283 week post-weaning (P < 0.01) with decreasing HC level, no dietary effects on growth 284 traits were observed in N-BtW pigs (Supplementary Table 1). 285

Table 3. Effect of decreasing hemicelluloses level in the maternal diet on the

287 performance of piglets

			² Contrasts				
	T12 -T13	T11	Т9	T7 -T8	SEM	L	Q
Body measurements at birth, cm							
Crown-to-rump length	28.7	28.9	28.8	28.4	0.53	0.60	0.56
Body circumference	25.5	25.6	25.8	25.3	0.49	0.81	0.57
Body mass index, kg/m ²	19.2	18.8	19.5	18.7	0.55	0.78	0.73
Ponderal index, kg/m ³	67.2	65.1	67.9	66.3	2.03	0.99	0.88
Body weight, kg							
At birth	1.61	1.60	1.63	1.52	0.083	0.55	0.50
5 days post-farrowing	2.38	2.37	2.45	2.22	0.126	0.49	0.36
16 days post-farrowing	5.36	5.28	5.25	4.94	0.272	0.29	0.65
Weaning	7.69	7.54	7.26	7.36	0.348	0.41	0.71
1 week post-weaning	7.82	7.69	7.42	7.48	0.371	0.55	0.92
2 week post-weaning	8.93	9.17	8.71	8.93	0.453	0.82	0.99
ADG, g/day							
Birth to 5 days post-farrowing	154	154	160	137	12.6	0.44	0.34
Birth to 16 days post-farrowing	235	230	225	212	13.9	0.25	0.74
Birth to weaning	237	232	222	222	11.5	0.29	0.82
Weaning to 2 weeks post-weaning	86	116	103	113	17.3	0.38	0.52
1 week to 2 weeks post-weaning	172	194	184	207	20.8	0.31	0.98
Birth-2 week post-weaning	185	191	180	184	9.9	0.73	0.90
Feed intake, g/piglet							
1 week pre-weaning	182	186	157	189	24.5	0.95	0.55
1 week post-weaning	753	883	760	786	121.0	0.96	0.65
2 weeks post-weaning	1428	1638	1436	1600	144.0	0.63	0.87
Post-weaning diarrhoea,%							
1 week post-weaning	26.1	29.3	27.0	29.6	2.47	0.47	0.77
2 weeks post-weaning	17.4	17.2	12.8	22.2	2.82	0.44	0.05
Days with diarrhoea, days							
1 week post-weaning	1.89	2.09	1.85	2.10	0.171	0.61	0.90
2 weeks post-weaning	1.45	1.40	1.11	1.80	0.172	0.34	0.02

 1 1

containing 11% of hemicelluloses; T9 = Sow's lactation diet containing 9% of hemicelluloses; T7 T8 =

290 Sow's lactation diet containing 78% of hemicelluloses.

291 ²Contrasts: L = Linear; Q = Quadratic.

Table 4. Effect of decreasing hemicelluloses level in maternal diet on the

293 performance of low birthweight piglets

	1	Dietary Tre	eatments			² Cont	rasts
	T12 T13	T11	Т9	17 -T8	SEM	L	Q
Number of piglets, <i>n</i>	25	23	15	20			
Body measurements at birth, cm							
Crown-to-rump length	25.0	25.4	25.5	26.0	0.56	0.22	0.97
Body circumference	22.3	21.8	21.9	22.3	0.42	0.98	0.16
Body mass index, kg/m ²	16.6	15.5	16.1	15.8	0.65	0.51	0.43
Ponderal index, kg/m³	66.6	61.4	63.6	61.7	3.22	0.37	0.54
Body weight, kg							
At birth	1.04	1.01	1.04	1.06	0.047	0.64	0.46
5 days post-farrowing	1.61	1.59	1.58	1.54	0.095	0.53	0.86
16 days post-farrowing	3.94	3.78	3.59	3.85	0.287	0.70	0.39
Weaning	5.86	5.73	5.42	6.55	0.468	0.38	0.12
1 week post-weaning	5.92	5.96	5.56	6.95	0.498	0.20	0.12
2 week post-weaning	6.55	6.66	6.43	8.35	0.545	0.02	0.06
ADG, g/day							
Birth to 5 days post-farrowing	113	118	104	91	14.1	0.14	0.43
Birth to16 days post-farrowing	181	173	158	173	16.7	0.57	0.41
Birth to weaning	192	184	171	201	16.3	0.83	0.18
Weaning to 2 weeks post-weaning	50	62	74	113	27.0	0.09	0.56
1 week to 2 weeks post-weaning	91	103	125	187	27.0	0.01	0.25
Birth to 2 weeks post-weaning	141	143	135	177	11.5	0.04	0.05
Post-weaning diarrhoea,%							
1 week post-weaning	19.8	34.1	16.4	20.8	10.50	0.69	0.50
2 weeks post-weaning	36.4	16.7	6.5	5.2	8.31	<0.01	0.35
Days in diarrhoea, days							
1 week post-weaning	1.66	2.26	1.32	1.63	0.502	0.56	0.71
2 weeks post-weaning	2.36	1.22	0.55	0.87	0.512	<0.01	0.07

294 1742 T13 = Sow's lactation diet containing 42 13% of hemicelluloses; T11 = Sow's lactation diet

containing 11% of hemicelluloses; T9 = Sow's lactation diet containing 9% of hemicelluloses; T7 T8 =

Sow's lactation diet containing 7 8% hemicellulose.

297 ²Contrasts: L = Linear; Q = Quadratic.

Throughout lactation, no dietary treatment and sampling day interaction was found 299 (data not shown). At days 3 and 17 of lactation, DM, ash, protein and somatic cell 300 count, as well as milk yield estimated from farrowing to day 3 and from day 4 to day 17 301 of lactation, were similar among dietary treatments (Table 5). With a decreasing HC 302 level, milk lactose content linearly decreased (P < 0.01). Regarding mineral levels in 303 the sow milk, calcium, phosphorus, sodium, magnesium and zinc contents remained 304 similar among experimental treatments, whereas the copper content linearly increased 305 (P = 0.02) with decreasing HC content in the maternal diet. Excluding the linear 306 increase (P = 0.04) in the threonine level and the quadratic increase (P = 0.04) in the 307 monounsaturated fatty acid portion, decreasing HC level in the maternal diet had no 308 impact on the amino acid and fatty acid profiles. Regardless of the dietary treatments, 309 somatic cell counts did not differ between the sampling days. However, the sampling 310 day influenced protein, mineral and lactose contents, as well as milk yield. Between 311 days 3 and 17 of lactation, protein, phosphorus, potassium and zinc contents 312 decreased ($P \le 0.05$), whereas lactose and calcium contents and milk yield increased 313 $(P \leq 0.05)$. Furthermore, histidine, leucine, isoleucine, phenylalanine, threonine, 314 tryptophan, tyrosine, valine, alanine, aspartic acid and serine decreased ($P \le 0.05$), 315 whereas glutamate and proline increased ($P \le 0.05$) between days 3 and 17. The fatty 316 acid profile in milk changed during lactation. Monounsaturated and polyunsaturated 317 fatty acid portions decreased ($P \le 0.05$) and saturated fatty acid content increased (P318 \leq 0.05) from day 3 to day 17. More precisely, the portions of C18:0, C18:1n-9, C18:2n-319 6, C18:3n-6, C18:3n-3, C20:4n-6, C20:5n-3 and C22:5n-3 decreased ($P \le 0.05$), 320 whereas C16:0 level increased ($P \le 0.05$) between days 3 and 17. 321

322 Table 5. Effect of decreasing hemicellulose level in sow's lactation diet on gross composition, mineral content, amino acid

323 profile and fatty acid profile of milk

	¹ C	Dietary T	reatmer	nts		² Cont	rasts	³ Stage	of lactation		
Item	T12 T13	T11	Т9	T7 T8	SEM	L	Q	d3	d17	SEM	<i>P</i> -value
Milk yield, kg/day	9.66	9.90	9.39	8.80	0.76	0.81	0.76	7.03	11.85	0.41	<0.01
Gross chemical composition											
Dry matter, %	19.5	20.7	19.9	20.6	0.60	0.25	0.49	20.7	19.7	0.40	0.06
Total protein, %	5.86	5.82	5.84	6.07	0.153	0.48	0.34	6.40	5.40	0.091	<0.01
Fat, %	7.50	8.65	8.07	8.69	0.533	0.15	0.38	8.50	7.96	0.364	0.27
Lactose, %	5.17	4.99	4.92	4.77	0.110	0.01	0.76	4.56	5.37	0.068	<0.01
Ash,%	0.86	0.86	0.88	0.85	0.150	0.29	0.89	0.89	0.83	0.098	<0.01
Somatic cells, log 10 ³ cells/ml	6.99	6.92	7.40	7.71	0.325	0.18	0.41	7.40	7.11	0.248	0.93
Gross energy, MJ/kg	5.14	5.70	5.43	5.70	0.230	0.10	0.30	5.65	5.34	0.162	0.72
Minerals											
Calcium, g/kg	1.91	1.98	2.02	1.99	0.051	0.97	0.94	1.88	2.07	0.033	<0.01
Phosphorus, g/kg	1.57	1.58	1.57	1.53	0.026	0.09	0.63	1.61	1.52	0.017	<0.01
Potassium, g/kg	1.11	1.10	1.11	1.05	0.028	0.07	0.24	1.29	0.90	0.019	<0.01
Sodium, g/kg	0.37	0.35	0.35	0.34	0.016	0.21	0.89	0.36	0.34	0.011	0.93
Magnesium, g/kg	0.10	0.11	0.11	0.11	0.003	0.42	0.09	0.11	0.11	0.002	0.57
Copper, mg/kg	1.37	1.45	1.51	1.76	0.135	0.02	0.28	1.68	1.37	0.085	0.67
Zinc, mg/kg	6.04	6.64	6.02	5.44	0.363	0.16	0.07	6.38	5.69	0.213	<0.01
Amino acids, % of total protein											
Alanine	3.28	3.29	3.33	3.35	0.025	0.21	0.46	3.41	3.21	0.017	<0.01
Arginine	4.57	4.62	4.68	4.67	0.029	0.13	0.99	4.72	4.55	0.020	<0.01
Aspartic acid	7.70	7.68	7.75	7.74	0.035	0.78	0.29	7.83	7.61	0.025	<0.01
Cysteine	1.40	1.39	1.39	1.42	0.015	0.15	0.10	1.44	1.36	0.010	<0.01
Glutamate	17.8	17.6	17.8	17.6	0.16	0.29	0.99	17.5	17.9	0.11	<0.01
Glycine	2.98	3.02	3.12	3.05	0.030	0.10	0.40	3.06	3.03	0.019	0.15

Histidine	2.53	2.53	2.53	2.56	0.015	0.85	0.31	2.56	2.51	0.009	<0.01
Isoleucine	3.85	3.80	3.80	3.83	0.039	0.36	0.53	3.84	3.80	0.022	0.05
Leucine	8.03	8.12	8.02	8.15	0.049	0.74	0.83	8.18	7.99	0.030	<0.01
Lysine	6.86	6.79	6.82	6.86	0.049	0.61	0.36	6.85	6.82	0.029	0.22
Methionine	1.74	1.72	1.71	1.71	0.014	0.10	0.68	1.72	1.72	0.008	0.52
Phenylalanine	3.86	3.85	3.87	3.92	0.026	0.17	0.13	3.92	3.83	0.017	<0.01
Proline	10.2	10.3	10.4	10.2	0.11	0.37	0.15	10.1	10.5	0.07	<0.01
Serine	4.70	4.66	4.73	4.76	0.047	0.15	0.39	4.75	4.67	0.030	0.02
Threonine	3.88	3.88	3.90	3.98	0.036	0.04	0.19	3.99	3.83	0.023	<0.01
Tryptophan	1.18	1.18	1.21	1.20	0.017	0.17	0.94	1.23	1.15	0.011	<0.01
Tyrosine	4.02	3.97	3.99	4.05	0.050	0.43	0.20	4.05	3.96	0.028	<0.01
Valine	5.16	5.21	5.20	5.26	0.039	0.17	0.98	5.30	5.12	0.025	<0.01
Fatty acids, % of total fatty acids											
C16:0	27.2	27.4	26.4	27.8	0.70	0.52	0.38	24.9	29.5	0.48	<0.01
C18:0	4.29	4.43	4.33	4.41	0.143	0.70	0.70	4.78	3.95	0.089	<0.01
C18:1n-9	35.3	36.1	35.8	34.8	0.83	0.51	0.38	37.2	33.8	0.57	<0.01
C18:2n-6	11.45	9.63	12.08	11.74	0.412	0.09	0.09	12.20	10.30	0.245	<0.01
C18:3n-6	0.14	0.12	0.15	0.13	0.012	0.93	0.87	0.20	0.08	0.008	<0.01
C18:3n-3	1.08	1.12	1.16	1.32	0.057	0.06	0.53	1.25	1.09	0.036	<0.01
C20:3n-3	0.11	0.11	0.11	0.09	0.010	0.76	0.48	0.11	0.10	0.006	0.06
C20:4n-6	0.52	0.50	0.55	0.55	0.022	0.16	0.53	0.65	0.41	0.014	<0.01
C20:5n-3	0.09	0.09	0.08	0.08	0.006	0.55	0.74	0.09	0.07	0.003	<0.01
C22:5n-3	0.23	0.22	0.21	0.22	0.018	0.77	0.61	0.26	0.18	0.010	<0.01
n-34	1.75	1.59	1.50	1.48	0.082	0.19	0.60	1.72	1.44	0.051	<0.01
<i>n</i> -6 ⁵	12.1	10.3	12.8	12.4	0.43	0.09	0.10	13.0	10.8	0.26	<0.01
Saturated	36.1	36.4	35.3	37.1	0.76	0.57	0.40	33.8	38.6	0.52	<0.01
Mono-unsaturated	49.1	50.8	49.5	48.1	0.61	0.71	0.04	50.4	48.3	0.42	<0.01
Poly-unsaturated	14.8	12.8	15.2	14.9	0.53	0.22	0.14	15.7	13.1	0.32	<0.01

324 ¹11² T13 = Sow's lactation diet containing <u>42</u> 13% of hemicelluloses; T11 = Sow's lactation diet containing 11% of hemicelluloses; T9 = Sow's lactation diet

325 containing 9% of hemicelluloses; ∓ 7 T8 = Sow's lactation diet containing 7 8% of hemicelluloses.

- 326 ²Contrasts: L = Linear; Q = Quadratic.
- 3 Days: d3 = Day 3 of lactation; d17 = Day 17 of lactation;
- ⁴*n*-3: sum of C18:3n-3, C20:3n-3, C20:5n-3, C22:5n-3.
- ⁵*n*-6: sum of C18:2n-6, C18:3n-6 and C20:4n-6.

330 Volatile fatty acid concentrations in milk

The VFA concentration and the proportion of butyrate linearly increased (P < 0.01; Table 6) with decreased HC content in the maternal diet, resulting in an increased in total VFA by 25% and butyrate proportion by 60%. Regardless of the dietary treatment, total VFA concentration decreased ($P \le 0.05$) by 71% between days 3 and 17. The proportion of methanoate increased (P < 0.01), and the proportion of acetate decreased (P < 0.01) between days 3 and 17, whereas the levels of propionate, isobutyrate, butyrate and isovalerate remained unchanged.

338 Table 6. Effect of decreasing hemicellulose levels in sow's lactation diet on the volatile fatty acid profile of milk

		¹ Dietary	Treatmen	ts		² Contrasts		³ Stage of lactation			
Item	T12 T13	T11	Т9	T7 T8	SEM	L	Q	d3	d17	SEM	<i>P</i> -value
Total volatile fatty acids, mmol/kg	3.07	3.58	3.60	3.86	0.28	0.03	0.60	4.12	2.94	0.19	<0.01
Proportion of individual VFA, %											
Methanoate	9.41	9.50	9.38	9.93	0.287	0.94	0.28	9.16	9.95	0.187	<0.01
Acetate	88.90	89.00	88.90	88.30	0.353	0.31	0.17	89.21	88.36	0.220	<0.01
Propionate	0.30	0.30	0.25	0.20	0.041	0.19	0.84	0.25	0.28	0.026	0.29
Isobutyrate	0.04	0.04	0.05	0.03	0.007	0.86	0.79	0.04	0.05	0.004	0.17
Butyrate	0.53	0.60	0.75	0.86	0.153	<0.01	0.64	0.68	0.69	0.104	0.29
Isovalerate	0.76	0.55	0.57	0.57	0.080	0.80	0.21	0.61	0.61	0.043	0.81

339 ¹112 T13 = Sow's lactation diet containing 42 13% of hemicelluloses; T11 = Sow's lactation diet containing 11% of hemicelluloses; T9 = Sow's lactation diet

340 containing 9% of hemicelluloses; ∓ 7 T8 = Sow's lactation diet containing 7% of hemicelluloses.

341 ²Contrasts: L = Linear; Q = Quadratic

 3 d3 = Day 3 of lactation; d17 = Day 17 of lactation

343 Discussion

344 Effect of decreasing the level of hemicelluloses on sows' performance

Excluding fibre intake, the sow's performances were not affected by dietary HC. As 345 expected, due to similar feed intake during the pre-farrowing and lactation periods, 346 decreasing the level of HC also reduced the intake of the low-molecular-weight DF and 347 SDF fractions. Renteria-Flores et al. (2008) found that increasing the SDF level in the 348 349 gestation diet increased BW loss during lactation. However, the adverse effects may be related to the inclusion level of SDF. Similar to the present study, Li et al. (2019b) 350 found no effect on sow's performance when the SDF level was decreased from 38.7 351 g/kg to 17.7 g/kg in the gestation diet during the whole gestation period Shang et al. 352 (2021) found no effect either on sow's BW or backfat thickness at farrowing and 353 weaning when the dietary SDF level was decreased from 40.6 g/kg to 13.9 g/kg in the 354 late gestation and from 27.2 g/kg to 14.3 g/kg during lactation. In addition, considerably 355 high SDF intake can negatively affect litter performance. Indeed, Liu et al. (2020) 356 reported that from day 90 of gestation to farrowing, a daily intake of 215 g of SDF (SDF: 357 45.7 g/kg as fed), compared with 138 g/day (29.7 g/kg as fed) and 96 g/day (17.8 g/kg), 358 decreases the number of piglets and litter weight at weaning. In the present study, 359 sows received between 133 and 83 g/day of SDF according to the diets, from day 110 360 of gestation to farrowing. This setup can explain the lack of differences in these traits 361 between the dietary treatments. Therefore, compared to the study of Liu et al. (2020), 362 the SDF intake during this period for the four treatment groups was not sufficiently 363 elevated to negatively impact litter performances. 364

365 Effect of decreasing hemicelluloses levels on milk composition and milk VFA 366 profile

Milk yield and composition play a crucial role in the growth of suckling piglets to reach 367 an adequate weaning weight. In the present study, decreasing the level of HC in the 368 maternal diet affected milk composition but did not affect milk yield. Furthermore, 369 lactose content decreased, whereas copper and threonine proportions increased with 370 decreased HC level. A previous study showed that glucose, glycerol and other glucose 371 precursors play an important role in the synthesis of lactose in sow's milk (Boyd et al., 372 1995). Houdijk et al. (2002) reported that the fermentation of SDF occurs already at 373 the end of the ileum. As decreasing the level of HC also decreased the intake of SDF, 374 one can hypothesize that lowering the HC supply reduced the absorbed HC 375 fermentation products available for lactose synthesis., the fermentability of organic 376 matter in the small intestine might have decreased. Moreover, a recent study showed 377 that a 3:1 ratio of IDF to SDF increases the fermentability of organic matter in the small 378 intestine (Hoogeveen et al., 2021). Therefore, a higher level of HC could be related to 379 the higher fermentability of the organic matter in the small intestine. Given Moreover, 380 due to the osmotic power of lactose (Costa et al., 2019), milk yield may drop together 381 with lactose as the HC level decreases (Costa et al., 2019). Surprisingly, milk yield only 382 decreased numerically, and this result could be due to the differences in lactose 383 concentration between the experimental groups, which were not sufficiently large to 384 affect milk yield. A further interest in the present study is the linear increase in copper 385 in milk with a decreased HC level. Copper is an essential microelement for animals, 386 with many biological functions, including iron metabolism, immunity, protection from 387 oxidative stress and improvement in the activity of digestive enzymes (Huang et al., 388

2015). The milk concentration of copper is affected by the source of the micromineral 389 (Peters et al., 2010). However, as the same micromineral source was used among the 390 four dietary treatments, the mechanism underlying the increase in copper 391 concentration remains unclear. Similarly, with decreased HC levels in the diet, the 392 proportion of threonine in the milk increased. This effect remains unclear, as the 393 calculated digestible threonine levels were similar between the ± 12 T13 and ± 7 T8 394 diets. Besides a similar DF content, hypothetically, decreasing the HC level using 395 several DF sources may affect the fermentation patterns in the gut, namely, the 396 concentration and proportion of VFA. As VFA can be absorbed, transported through 397 the blood and finally reach the mammary glands, modifications in the milk composition 398 are expected (Tian et al., 2020). Decrease in HC level increased total VFA 399 concentration and butyrate proportion in milk. Zhao et al. (2019) showed a positive 400 correlation between VFA concentration in pig's ileum and decreased HC level. Given 401 that sows can ferment DFs better than growing pigs, a similar phenomenon may have 402 occurred in the ileum of sows fed with a low HC level (Noblet and Le Goff, 2001). 403 Furthermore, this effect on VFA in milk may also be due to differences in the intake of 404 other DF fractions. As previously mentioned, decreasing HC level concomitantly 405 increased ADF intake and decreased SDF intake. A positive correlation was reported 406 between the ADF level in pig's diet and butyrate concentration in the faeces (Zhao et 407 al., 2019). In the present study, hypothetically, increased ADF intake in sows fed with 408 decreasing level of HC might have increased the butyrate proportion in the faeces and 409 then in the milk. Compared with IDF, SDF is rapidly fermented by bacteria, thereby 410 enhancing the production of VFA (Jha and Berrocoso, 2015). Therefore, with 411 decreased SDF intake, VFA production should be lowered. However, the present study 412 showed that this concept was not evident and confirmed the importance of the source 413

of DF, as reported by some authors (Theil et al., 2014). Therefore, to understand the
effects of DF on milk composition, different fractions of DF, including HC and ADF
contents, must be considered.

417 Effects of the lactation diet on piglets' performance

In the present study, modifying the level of HC in the maternal diet did not enhance 418 litter performance. This result is consistent with the results of Loisel et al. (2013), which 419 showed that modifying the maternal diet is easier to positively affect the performances 420 of L-BtW piglets than the performance of the litter overall. Therefore, decreasing the 421 HC level improved post-weaning performance and reduced the occurrence of 422 diarrhoea in the L-BtW piglets. By contrast, why the performance of N-BtW was 423 unaffected by the HC level even though the occurrence of diarrhoea increased in this 424 group remains unclear. The L-BtW piglets usually exhibit poor performances, such as 425 a high mortality rate and low ADG, which represents high economic costs for farmers 426 due to reduced slaughter weight and increased occupancy of the stables (López-Vergé 427 et al., 2018). Girard et al. (2021) highlighted the importance of early-life interventions 428 to improve the post-weaning development and health of this sub-population of piglets. 429 Therefore, t In the present study, the beneficial effects observed in L-BtW piglets during 430 post-weaning period like the improved growth performance and the lower incidence of 431 diarrhoea may be related to the combination of an increased relative abundance of 432 butyrate, threonine and copper and to an increased concentration of total VFA in milk. 433 Indeed Given that piglets are highly susceptible to intestinal bacterial disorders during 434 the post-weaning period, butyrate, due to its recognised role in gut health, could have 435 been useful in increasing gut impermeability, alleviating diarrhoea in L-BtW piglets 436 during the second week post-weaning (Feng et al., 2018). In addition, given that piglets 437

are highly susceptible to intestinal bacterial disorders during the post-weaning period, 438 increasing threonine and copper proportions in the milk in the pre-weaning period can 439 help accelerate the gut maturation of those piglets (Lalles et al., 2009). Threonine plays 440 a critical role in the regulation of intestinal mucosal integrity, as it is required for the 441 production of mucins and immunoglobulins, improving the physical protection from the 442 attachment of microbes to the mucosal surface (Van Klinken et al., 1995). By contrast, 443 copper can help against pathogenic bacteria because of its bacteriostatic properties, 444 which affect the community structure of microorganisms in the caecum and colon 445 (Højberg et al., 2005). A lower relative abundance of Alistipes, Lachnospiraceae, 446 Ruminococcaceae and Prevotellaceae has been reported in the colon and ileum of L-447 BtW piglets compared with N-BtW piglets (Li et al., 2019a). These genera enhance gut 448 health and immune functions in the host (Den Besten et al., 2013). Given that, 449 colostrum and mature milk are key components in shaping piglet microbiota (Trevisi et 450 al., 2021), the modification of milk composition induced by decreased HC level in the 451 sow diet might have changed the gut microbiota of L-BtW piglets and improved their 452 health and growth. 453

454 Effect of lactation stage on milk composition

Sow's milk composition is strongly affected by changes throughout the lactation period. Colostrum (0–24 h after parturition) and Transitional milk (48–72 h after parturition) contain higher amounts of lipids, protein and dry matter compared with mature milk (from day 10 of lactation) (Csapó et al., 1996). In the present study, the passage from transitional milk to mature milk was characterised by a decrease in protein and ash contents and an increase in lactose content. Nevertheless, the contents of fat, dry matter and gross energy decreased only numerically from day 3 to day 17. Indeed, in

the present experiment, the lack of statistical differences on those traits is in 462 disagreement with several studies (Csapó et al., 1996; Theil et al., 2014), could be due 463 to the where differences between the sampling days were reported, which were not 464 large enough to affect dry matter, fat and gross energy contents in the sow's milk. This 465 might be related to differences in sow genotypes, sow management and litter size 466 between the present study and the previous ones. Similarly, the decrease in amino 467 acid proportion follows the same trend as protein content, except for glycine, lysine 468 and methionine, which remained stable over lactation, and for glutamate and proline, 469 which increased from day 3 to day 17. Therefore, the high level of amino acids in 470 471 transitional milk reflects the protein level, mainly because of the high content of immunoglobulins (Klobasa et al., 1987). The mineral content and fatty acid profile were 472 was also affected by the stage of lactation with an increase in the calcium level and a 473 decrease in the potassium and zinc levels from transitional milk to mature milk in 474 agreement with Csapó et al. (1996). Calcium and phosphorus play a key role in 475 improving piglet's growth (Hu et al., 2019 These changes are consistent with a previous 476 study (Csapó et al., 1996) that also reported an increase in the calcium level and a 477 decrease in the potassium and zinc levels from transitional milk to mature milk. The 478 present experiment also showed. Moreover, the phosphorus content decreased 479 between days 3 and 17. The reason for this decrease over lactation remains unclear 480 but might be related to a dilution effect, as it follows the numerical decrease in dry 481 matter. When expressed per kilogram of dry matter, the phosphorus concentration was 482 similar between days 3 and 17. Moreover, from transitional milk to mature milk, the 483 decrease in the proportion of mono- and polyunsaturated fatty acids and the increase 484 in the proportion of saturated fatty acids are related to changes in the proportion of 485 individual fatty acids. The increase in C16:0 proportion and decrease in the proportions 486

of C18:0, C18:1n-9, C18:2n-6, C18:3n-6, C18:3n-3, C20:4n-6 and C20:5n-3 observed
in the present study have already been described in a previous study (Hu et al., 2019).
Furthermore, Hu et al. (2019) reported a positive correlation between calcium and
C16:0 fatty acid.-Indeed, long-chain saturated fatty acids can form calcium salts in the
piglet's intestine, which can improve dry matter digestibility and enhance growth
performance (Kluge et al., 2006). However, the mechanism by which calcium affects
the synthesis of C16:0 remains unknown.

In conclusion, as shown in the present study, when the DF level is the same, feeding 494 lactating sows with a lower HC level can positively affect the milk composition and 495 offspring the development of L-BtW piglets. As HC content decreased, the growth 496 performance of the L-BtW piglets improved after weaning, and the occurrence of 497 diarrhoea decreased, particularly in the second week post-weaning. Moreover, it 498 increased the proportion of butyrate, copper and threonine and increased the VFA 499 concentration in the milk. Therefore, this study highlighted the importance of the 500 maternal diet in lactation to positively affect the development and health of L-BtW 501 piglets in the post-weaning period. 502

Ethics approval

The experiment was conducted in accordance with the Swiss Guidelines for Animal Welfare, and the Swiss Cantonal Committee for Animal Care and Use approved all procedures involving animals (approval number: 2019_25_FR).

Data accessibility

The data that support the findings of this study are publicly available in Zenodo (<u>https://doi.org/10.5281/zenodo.5814624</u>).

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Author contributions

Francesco Palumbo and Marion Girard validated the data and carried out the main statistical analyses. Marion Girard and Giuseppe Bee conceived the study design and secured substantial funding. Francesco Palumbo and Marion Girard performed the animal experiment, recorded the data and collected and processed the milk samples. Marion Girard, Francesco Palumbo, Giuseppe Bee and Paolo Trevisi supervised analyses and drafted and critically reviewed the manuscript. All authors read and approved the final manuscript.

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Declaration of interest

The authors declare they have no conflict of interest relating to the content of this article.

Supplementary materials

The Supplementary Table S1 and the statistical codes used can be found in the Supplementary Materials.

References

Agroscope. 2018. Fütterungsempfehlungen und Nährwerttabellen für Schweine. LmZ, Zollikofen 242

Boyd, D. R., R. S. Kensinger, R. J. Harrell, and D. E. Bauman. 1995. Nutrient uptake and endocrine regulation of milk synthesis by mammary tissue of lactating sows. Journal of Animal Science 73(suppl_2):36-56/DOI: https://doi.org/10.2527/1995.73suppl_236x. Costa, A., N. Lopez-Villalobos, N. Sneddon, L. Shalloo, M. Franzoi, M. De Marchi, and M. Penasa. 2019. Invited review: Milk lactose—Current status and future challenges in dairy cattle. Journal of dairy science 102(7):5883-5898/DOI: https://doi.org/10.3168/jds.2018-15955.

Csapó, J., T. Martin, Z. Csapo-Kiss, and Z. Hazas. 1996. Protein, fats, vitamin and mineral concentrations in porcine colostrum and milk from parturition to 60 days. International Dairy Journal 6(8-9):881-902/DOI: https://doi.org/10.1016/0958-6946(95)00072-0.

Den Besten, G., K. Van Eunen, A. K. Groen, K. Venema, D.-J. Reijngoud, and B. M. Bakker. 2013. The role of short-chain fatty acids in the interplay between diet, gut microbiota, and host energy metabolism. Journal of lipid research 54(9):2325-2340/DOI: https://doi.org/10.1194/jlr.R036012.

Dourmad, J., M. Etienne, and J. Noblet. 2001. Measuring backfat depth in sows to optimize feeding strategy. INRA Productions Animales, 14: 41-50 (France).

Feng, W., Y. Wu, G. Chen, S. Fu, B. Li, B. Huang, D. Wang, W. Wang, and J. Liu. 2018. Sodium butyrate attenuates diarrhea in weaned piglets and promotes tight junction protein expression in colon in a GPR109A-dependent manner. Cellular Physiology and Biochemistry 47(4):1617-1629/DOI: https://doi.org/10.1159/000490981

Girard, M., M. Tretola, and G. Bee. 2021. A Single Dose of Synbiotics and Vitamins at Birth Affects Piglet Microbiota before Weaning and Modifies Post-Weaning Performance. Animals 11(1):84/DOI: https://doi.org/10.3390/ani11010084 Hales, J., V. A. Moustsen, M. B. F. Nielsen, and C. F. Hansen. 2013. Individual physical characteristics of neonatal piglets affect preweaning survival of piglets born in a noncrated system. Journal of Animal Science 91(10):4991-5003/DOI: https://doi.org/10.2527/jas.2012-5740

Hoogeveen, A. M., Moughan, P. J., Henare, S. J., Schulze, P., McNabb, W. C., & Montoya, C. A. (2021). Type of Dietary Fiber Is Associated with Changes in Ileal and Hindgut Microbial Communities in Growing Pigs and Influences In Vitro Ileal and Hindgut Fermentation. *The Journal of Nutrition*, *151*(10), 2976-2985/DOI: https://doi.org/10.1093/jn/nxab228

Højberg, O., N. Canibe, H. D. Poulsen, M. S. Hedemann, and B. B. Jensen. 2005. Influence of dietary zinc oxide and copper sulfate on the gastrointestinal ecosystem in newly weaned piglets. Applied and environmental microbiology 71(5):2267-2277/DOI: https://doi.org/10.1128/AEM.71.5.2267-2277.2005

Houdijk, J. G., Verstegen, M. W., Bosch, M. W., & van Laere, K. J. 2002. Dietary fructooligosaccharides and transgalactooligosaccharides can affect fermentation characteristics in gut contents and portal plasma of growing pigs. Livestock Production Science, 73(2-3), 175-184. https://doi.org/10.1016/S0301-6226(01)00250-0

Hu, P., H. Yang, B. Lv, D. Zhao, J. Wang, and W. Zhu. 2019. Dynamic changes of fatty acids and minerals in sow milk during lactation. Journal of animal physiology and animal nutrition 103(2):603-611/DOI: https://doi.org/10.1111/jpn.13040

Huang, L. Z., Ma, M. G., Ji, X. X., Choi, S. E., & Si, C. 2021. Recent developments and applications of hemicellulose from wheat straw: a review. *Frontiers in Bioengineering and Biotechnology*, *9*, 440/DOI: https://doi.org/10.3389/fbioe.2021.690773.

Huang, Y., M. Ashwell, R. Fry, K. Lloyd, W. Flowers, and J. Spears. 2015. Effect of dietary copper amount and source on copper metabolism and oxidative stress of weanling pigs in short-term feeding. Journal of Animal Science 93(6):2948-2955/DOI: https://doi.org/10.2527/jas.2014-8082.

Jha, R., and J. Berrocoso. 2015. Dietary fiber utilization and its effects on physiological functions and gut health of swine. Animal 9(9):1441-1452/DOI: https://doi.org/10.1017/S1751731115000919

Jiménez-Escrig, A., & Sánchez-Muniz, F. J. (2000). Dietary fibre from edible seaweeds: Chemical structure, physicochemical properties and effects on cholesterol metabolism. Nutrition research, 20(4), 585-598. https://doi.org/10.1016/S0271-5317(00)00149-4

Klobasa, F., E. Werhahn, and J. Butler. 1987. Composition of sow milk during lactation. Journal of animal science 64(5):1458-1466/DOI: https://doi.org 10.2527/jas1987.6451458x

Kluge, H., J. Broz, and K. Eder. 2006. Effect of benzoic acid on growth performance, nutrient digestibility, N balance, gastrointestinal microflora and parameters of microbial metabolism in piglets. Journal of Animal Physiology and Animal Nutrition 90(7-8):316-324/DOI: https://doi.org/10.1111/j.1439-0396.2005.00604.x

Kragten, S. A., M. Collomb, S. Dubois, and P. Stoll. 2014. Composition des acides gras dans l'alimentation animale–méthodes d'analyse. Rech. Agron. Suisse 5:330-337.

Lalles, J. P., P. Bosi, P. Janczyk, S. Koopmans, and D. Torrallardona. 2009. Impact of bioactive substances on the gastrointestinal tract and performance of weaned piglets: a review. Animal 3(12):1625-1643/ DOI: https://doi.org/10.1017/S175173110900398X

Lattimer, J. M., and M. D. Haub. 2010. Effects of dietary fiber and its components on metabolic health. Nutrients 2(12):1266-1289/DOI: https://doi.org/10.3390/nu2121266

Li, N., S. Huang, L. Jiang, Z. Dai, T. Li, D. Han, and J. Wang. 2019a. Characterization of the early life microbiota development and predominant Lactobacillus species at distinct gut segments of low-and normal-birth-weight piglets. Frontiers in microbiology 10:797/DOI: https://doi.org/10.3389/fmicb.2019.00797

Li, Y., L. Zhang, H. Liu, Y. Yang, J. He, M. Cao, M. Yang, W. Zhong, Y. Lin, and Y. Zhuo. 2019b. Effects of the ratio of insoluble fiber to soluble fiber in gestation diets on sow performance and offspring intestinal development. Animals 9(7):422/DOI: https://doi.org/10.3390/ani9070422

Liu, Y., N. Chen, D. Li, H. Li, Z. Fang, Y. Lin, S. Xu, B. Feng, Y. Zhuo, and D. Wu. 2020. Effects of dietary soluble or insoluble fiber intake in late gestation on litter performance, milk composition, immune function, and redox status of sows around parturition. Journal of Animal Science 98(10):skaa303/DOI: https://doi.org/10.1093%2Fjas%2Fskaa303

Loisel, F., Farmer, C., Ramaekers, P., & Quesnel, H. (2013). Effects of high fiber intake during late pregnancy on sow physiology, colostrum production, and piglet performance. *Journal of Animal Science*, *91*(11), 5269-5279/DOI: https://doi.org/10.2527/jas.2013-6526

López-Vergé, S., J. Gasa, M. Farré, J. Coma, J. Bonet, and D. Solà-Oriol. 2018. Potential risk factors related to pig body weight variability from birth to slaughter in commercial conditions. Translational Animal Science 2(4):383-395/DOI: https://doi.org/10.1093/tas/txy082 Montoya, C. A., Saigeman, S., Rutherfurd, S. M., & Moughan, P. J. 2016. The digestion of kiwifruit (Actinidia deliciosa) fibre and the effect of kiwifruit on the digestibility of other dietary nutrients. Food Chemistry, 197, 539-545.

Noblet, J., and G. Le Goff. 2001. Effect of dietary fibre on the energy value of feeds for pigs. Animal feed science and technology 90(1-2):35-52/DOI: https://doi.org/10.1016/S0377-8401(01)00195-X

Paßlack, N., W. Vahjen, and J. Zentek. 2015. Dietary inulin affects the intestinal microbiota in sows and their suckling piglets. BMC veterinary research 11(1):1-8/DOI: https://doi.org/10.1186/s12917-015-0351-7

Peters, J., D. Mahan, T. Wiseman, and N. Fastinger. 2010. Effect of dietary organic and inorganic micromineral source and level on sow body, liver, colostrum, mature milk, and progeny mineral compositions over six parities. Journal of animal science 88(2):626-637/DOI: https://doi.org/10.2527/jas.2009-1782

Renteria-Flores, J., L. J. Johnston, G. C. Shurson, and D. D. Gallaher. 2008. Effect of soluble and insoluble fiber on energy digestibility, nitrogen retention, and fiber digestibility of diets fed to gestating sows. Journal of Animal Science 86(10):2568-2575/-DOI: https://doi.org/10.2527/jas.2007-0375

Shang, Q., Liu, S., Liu, H., Mahfuz, S., & Piao, X. (2021). Impact of sugar beet pulp and wheat bran on serum biochemical profile, inflammatory responses and gut microbiota in sows during late gestation and lactation. Journal of Animal Science and Biotechnology, 12(1), 1-14. https://doi.org/10.1186/s40104-021-00573-3

Theil, P. K., C. Flummer, W. Hurley, N. B. Kristensen, R. Labouriau, and M. T. Sørensen. 2014. Mechanistic model to predict colostrum intake based on deuterium

oxide dilution technique data and impact of gestation and prefarrowing diets on piglet intake and sow yield of colostrum. Journal of Animal Science 92(12):5507-5519/DOI: https://doi.org/10.2527/jas.2014-7841

Tian, M., J. Chen, J. Liu, F. Chen, W. Guan, and S. Zhang. 2020. Dietary fiber and microbiota interaction regulates sow metabolism and reproductive performance. Animal Nutrition 6:397–403/DOI: https://doi.org/10.1016/j.aninu.2020.10.001

Trevisi, P., D. Luise, F. Correa, and P. Bosi. 2021. Timely Control of Gastrointestinal Eubiosis: A Strategic Pillar of Pig Health. Microorganisms 9(2):313/DOI: https://doi.org/10.3390/microorganisms9020313

Van der Peet-Schwering, C., J. Swinkels, and L. Den Hartog. 1998. Nutritional strategy and reproduction. The Lactating Sow. Wageningen Pers, Wageningen: 221-240.

Van Klinken, B., J. Dekker, H. Buller, and A. Einerhand. 1995. Mucin gene structure and expression: protection vs. adhesion. American Journal of Physiology-Gastrointestinal and Liver Physiology 269(5):G613-G627/ DOI: https://doi.org 10.1152/ajpgi.1995.269.5.G613

Van Soest, P. V., Robertson, J. B., and Lewis, B. A. 1991. Methods for dietary fiber, neutral detergent fiber, and nonstarch polysaccharides in relation to animal nutrition. Journal of dairy science, 74(10), 3583-3597. https://doi.org/10.3168/jds.S0022-0302(91)78551-2

Zhao, J., Y. Bai, S. Tao, G. Zhang, J. Wang, L. Liu, and S. Zhang. 2019. Fiber-rich foods affected gut bacterial community and short-chain fatty acids production in pig model. Journal of Functional Foods 57:266-274/DOI: https://doi.org/10.1016/j.jff.2019.04.00